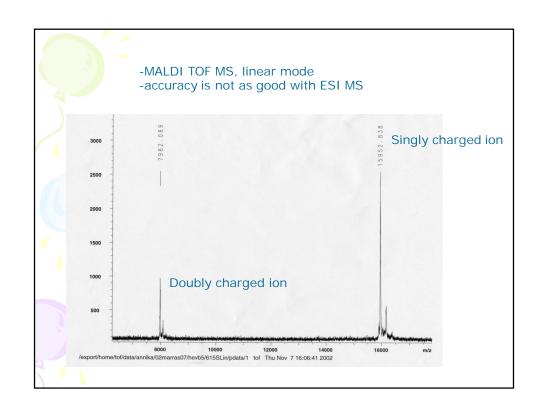
Protein MW determination and protein identification by mass spectrometry

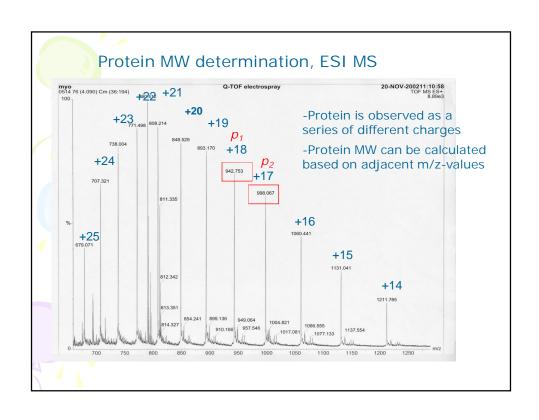
Tuula Nyman
Protein Chemistry Research Group
Institute of Biotechnology
tuula.nyman@helsinki.fi

Protein MW determination by MS (NOT= identification)

-for MW determination the protein needs to be in solution without salts and detergents

-usually proteins are first purified with RP chromatography before MW measurement





Protein MW calculation from ESI spectra

$$p=m/z$$

$$p_1 = (M_r + z_1)/z_1$$

$$p_2 = [M_r + (z_1-1)]/(z_1-1)$$

p= a peak in the mass spectrum m= total mass of an ion z= total charge $M_r=$ average mass of the protein

p=m/z $p_1=(M_r+z_1)/z_1$ $p_2=[M_r+(z_1-1)]/(z_1-1)$

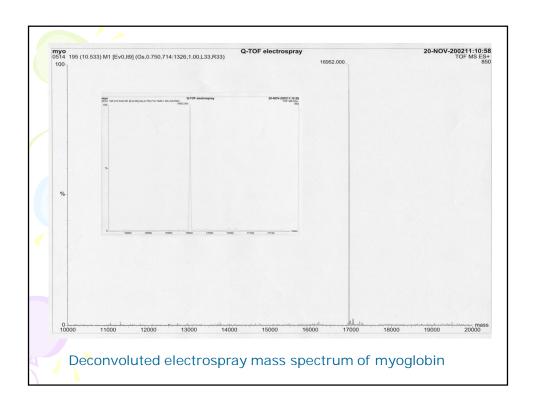
If p_1 =942.753 and p_2 =998.067

$$942.753 = (M_r + z_1)/z_1$$

 $942.753z_1 = Mr + z_1$
 $941.753z_1 = M_r$

998.067 = $(941.753z_1 + z_1 - 1)/z_1 - 1$ 998.067 $z_1 - 997.067 = 942.753z_1 - 1$ $(998.067 - 942.753)z_1 = 996.067$ $55.314z_1 = 996.067 = > z_1 = 18.0075$

 $M_r = 941.753z_1 = 16 951.6 \text{ Da}$



Protein identification by mass spectrometry

- •protein of interest is cleaved into peptides with a specific enzyme
- peptides are analyzed by MS (and MS/MS)

Protein identification methods:

- Peptide mass fingerprinting (PMF)
- •Identification based on MS/MS data from one or more peptides

Peptide mass fingerprint (PMF)

A mass spectrum of the peptide mixture resulting from the digestion of a protein by an enzyme, usually measured by MALDI-TOF

Identification based on peptide MW information only

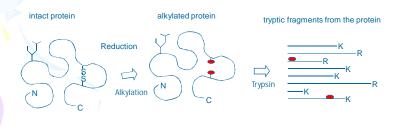
Database search engines create theoretical PMFs for all the proteins in the database and compare these to the measured PMF from protein X

Protein identification based on MS/MS data from one or more peptides

Database search engines create theoretical peptide fragmentation patterns for all the proteins in the database and compare these to the measured MS/MS data

Protein identification by MS

- Protein has to be digested into peptides
- •Disulphide bridges need to be reduced and alkylated before digestion



Protein digestion into peptides before MS analysis

Digestion can be done *in-solution* or *in-gel*The enzyme has to be as specific as possible

Trypsin is most commonly used enzyme:

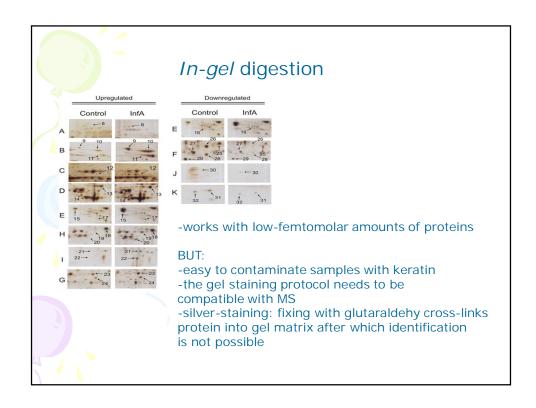
- -very specific, quite cheap
- -cleaves peptide bond after lysines and arginines
- -tryptic peptides are 'good' for MS analysis because they end up with basic amino acid
- -works for both *in-solution* and *in-gel* digestion

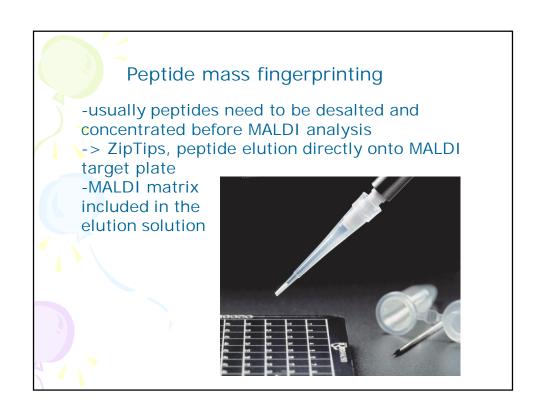
Other enzymes:

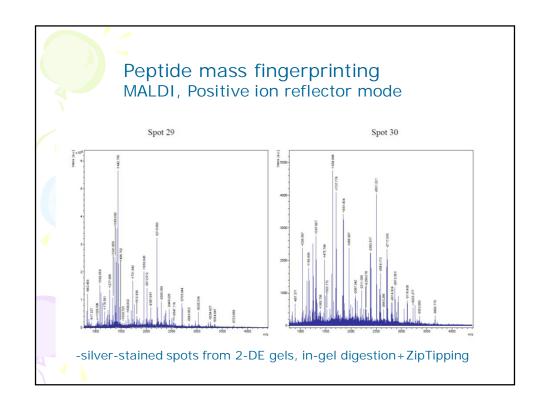
- -LysC, cleaves after lysines
- -LysN, cleaves before lysines
- -AspN, cleaves before aspartic acid residues
- V8 protease, cleaves peptide bonds exclusively on the carbonyl side of aspartate and glutamate residues
- -possible to do double-digestions

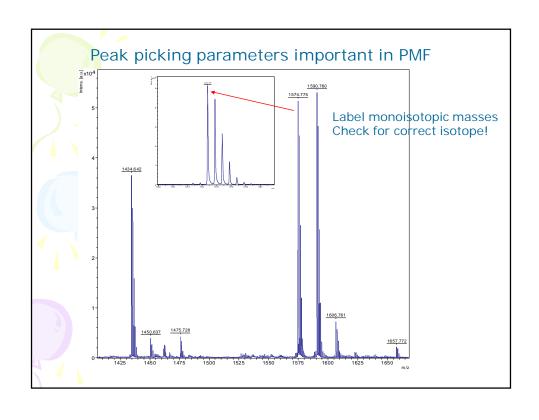
Chemical cleavage:

Cyanogen bromide, cuts after methionines







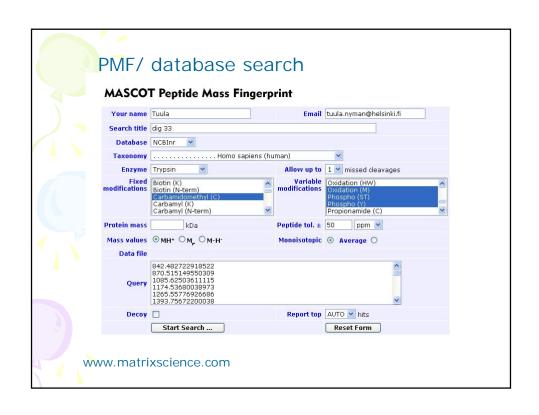


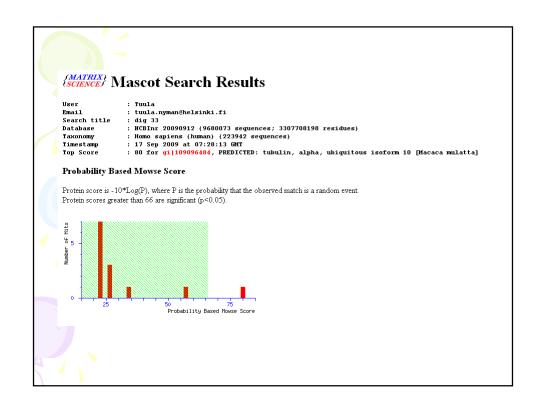
Publicly available search engines for PMF

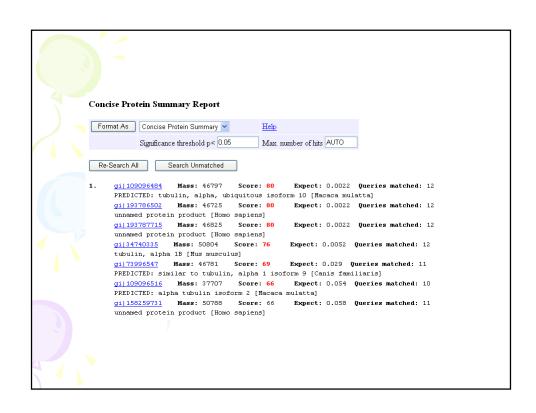
- Mascot
- ProteinProspector/ MS-Fit
- •PROWL/ ProFound
- Aldente

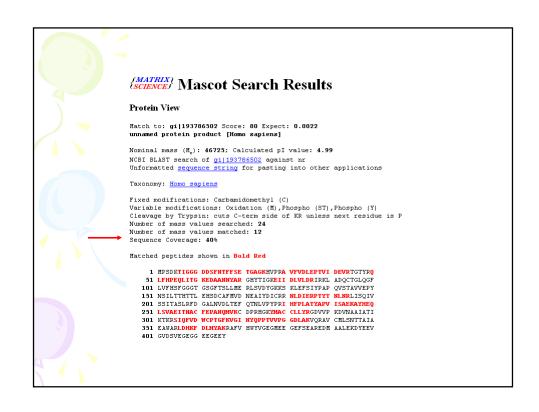
Databases

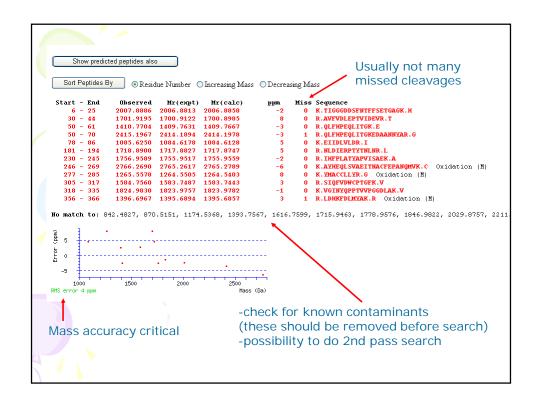
Database	Comment
EST	EST divisions of Genbank, (currently EST_human, EST_mouse, EST_others)
MSDB	Comprehensive, non-identical protein database
NCBInr	Comprehensive, non-identical protein database
SwissProt	High quality, curated protein database

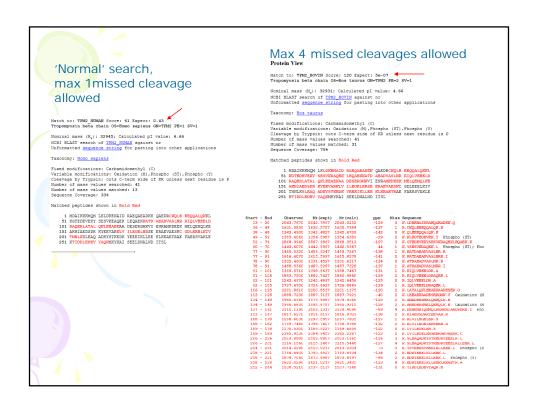










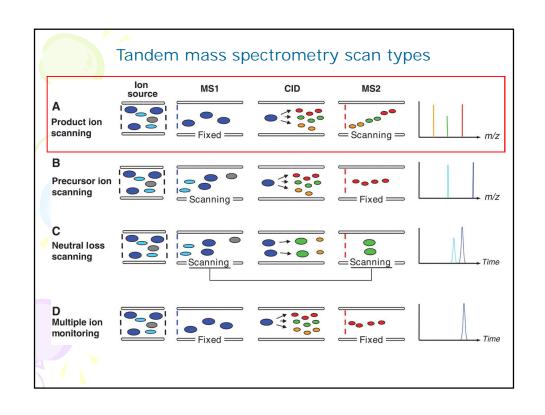


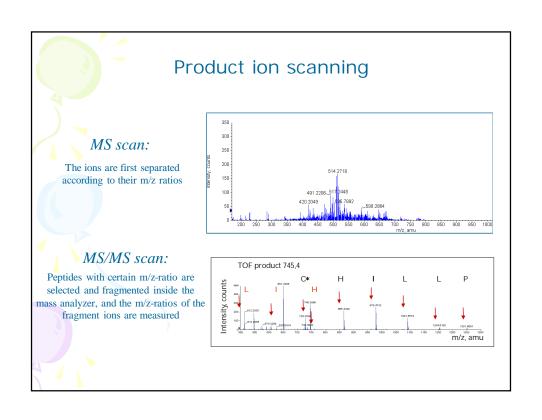
PEPTIDE MASS FINGERPRINTING

- 'quick and easy'
- requires
 - a very spesific enzyme
 - optimized digestion+ desalting protocols
 - internal/close external calibration of MALDI spectra
- works only for proteins which are already in the databases as protein sequences
- not suitable for complex protein mixtures

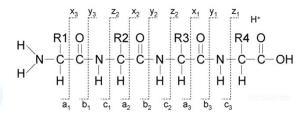
Protein identification based on MS/MS data from one or more peptides

- -MALDI-TOF/TOF or nanoLC-ESI-MS/MS analysis
- -suitable for (complex) protein mixtures, especially when combined with LC separation of peptides before MS/MS





Peptide Fragmentation Nomenclature

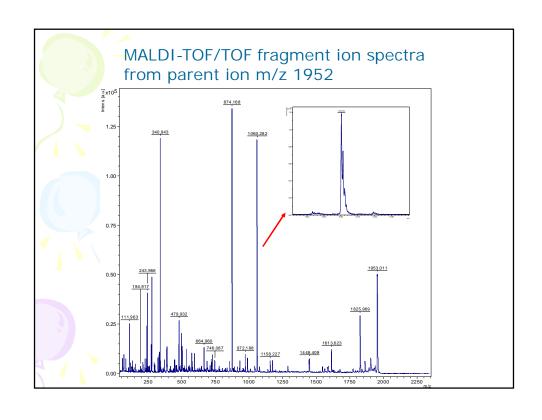


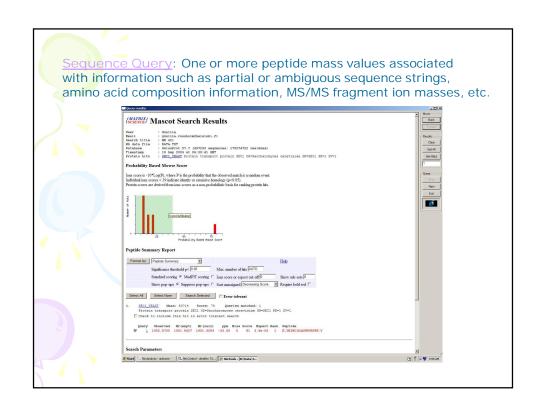
Peptides do not fragment sequentially, the fragmentation events are somewhat random.

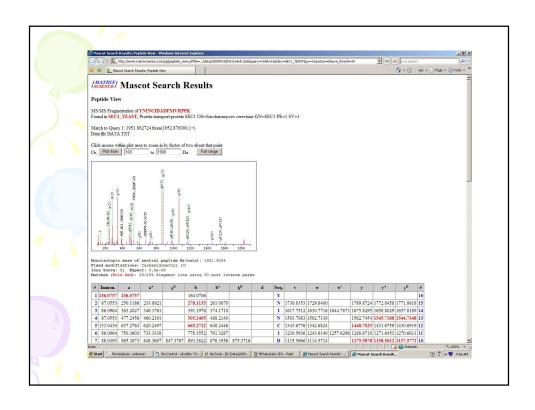
The most common peptide fragments observed in low energy collisions are a, b and y ions. The b ions appear to extend from the amino terminus (N-terminus), and y ions appear to extend from the carboxyl terminus (C-terminus).

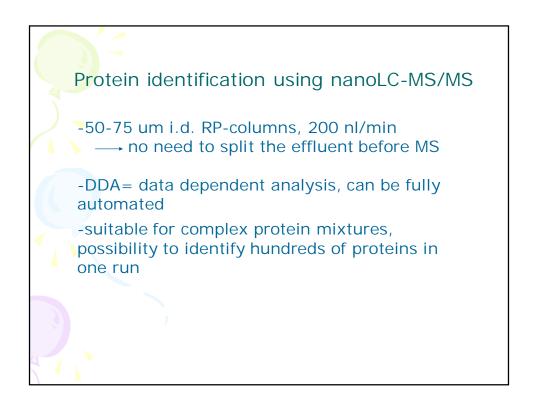
Protein identification with MALDI-TOF/TOF

- -first, PMF with MALDI-TOF
- -next, selected precursor ions can be fragmented (TOF/TOF analysis)
- MALDI produces singly charged parent ions
 → product ion spectra not as easy to interpret as in ESI-MS/MS
- -database search with both PMF and MS/MS information



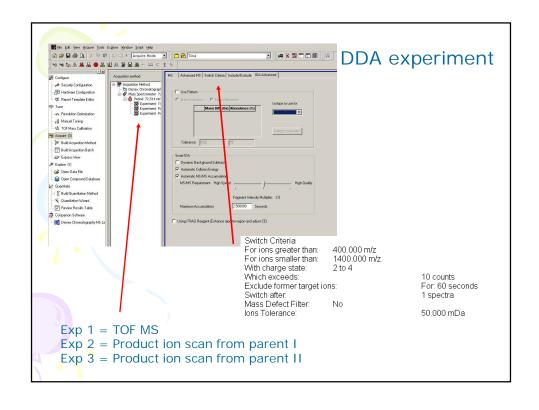


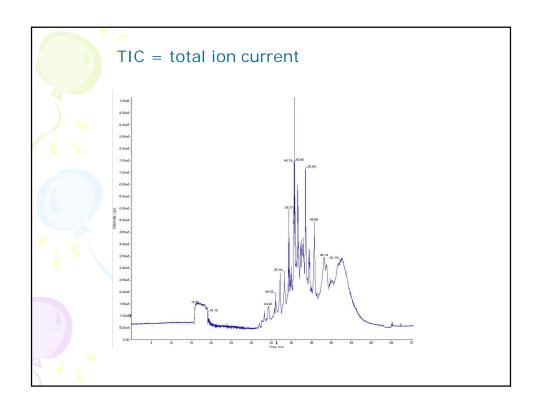


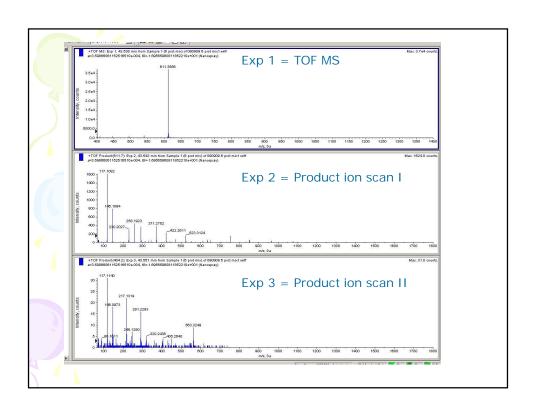


DDA = Data Dependent Acquisition (IDA = Information Dependent Acquisition)

- -fully automated experiment, first MS scan followed by two or more product ion scans
- -the acquisition software is set to choose certain types of ions for fragmentation and to use 'suitable' collision energy for this ion
- -in ESI tryptic peptides have usually 2-4 charges

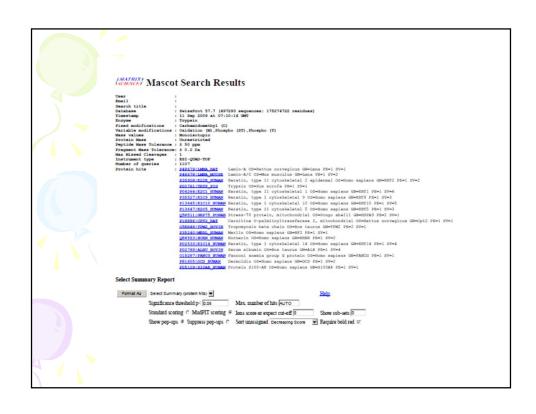




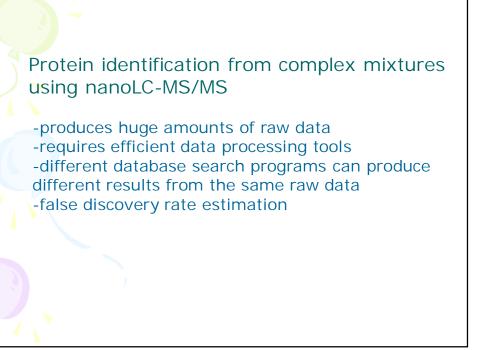


Search engines for (LC-)MS/MS data

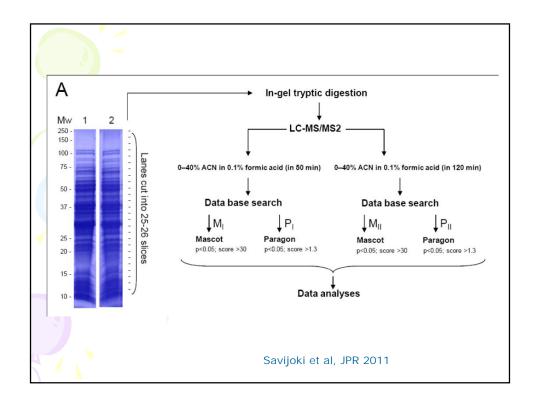
- -Mascot, Sequest, OMSSA etc
- -the programs take the fragment ion spectrum of a peptide as input and score it against theoretical fragmentation patterns constructed for peptides from the searched database.
- -in practise the user is often limited to use those search engines which accept the data format from the mass spec used
- -mzXML is a open data format for storage and exchange of mass spec data
- -raw, proprietary file formats from most vendors can be converted to the open mzXML format

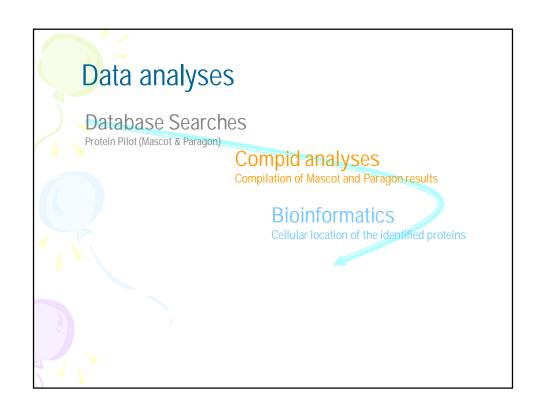


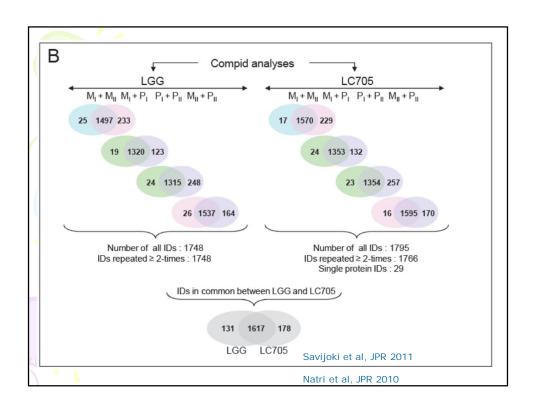




Comparative proteome cataloging of Lactobacillus rhamnosus strains GG and Lc705 -in-depth proteome analysis of two Lactobacillus rhamnosus strains, the well-known probiotic strain GG and the dairy strain Lc705 -GeLC-MS/MS: proteins are separated using SDS_PAGE and identified using nanoLC-MS/MS -to maximize the number of identifications, all data sets were searched against the target databases using two search engines, Mascot and Paragon



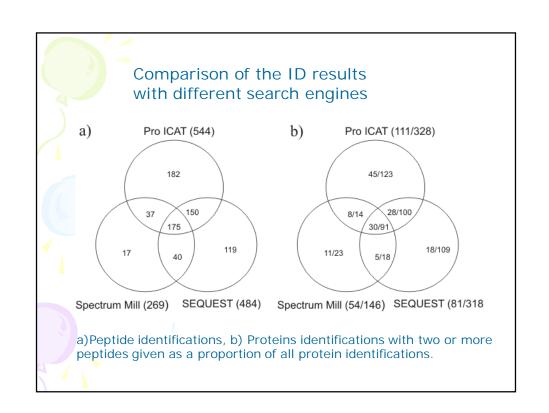




Comparison of protein ID results from the same raw data with different search engines

• ProICAT SP2
• Spectrum Mill
• Sequest

Moulder et al, Proteomics. 2005;5:2748-60.



False discovery rate (FDR) estimation

FDR = FP/(TP+FP)

TP= true positive matches FP = false positive matches

Previously: search the raw data first against the 'normal' database and then against the decoy database -> calculate FDR based on these

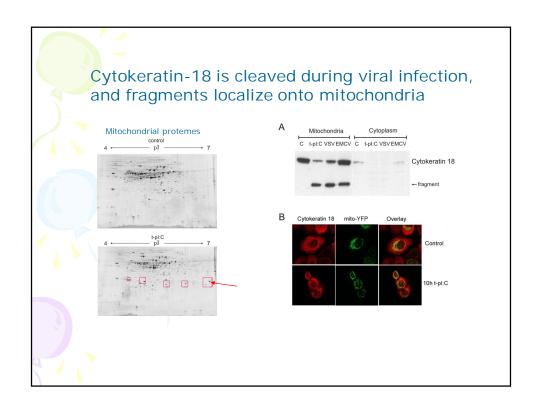
NOW the preferred method:

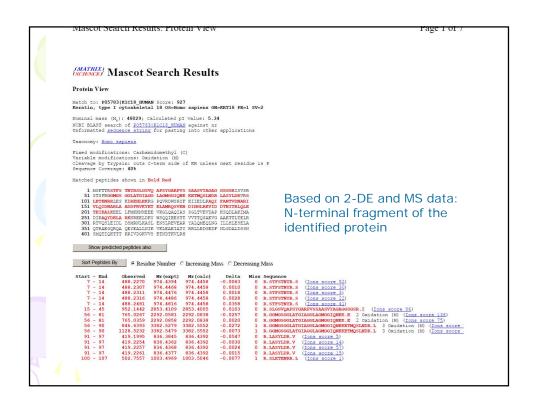
One search against a database in which the target and decoy sequences have been concatenated. This means that you will only record a false positive when a match from the decoy sequences is better than any match from the target sequences.

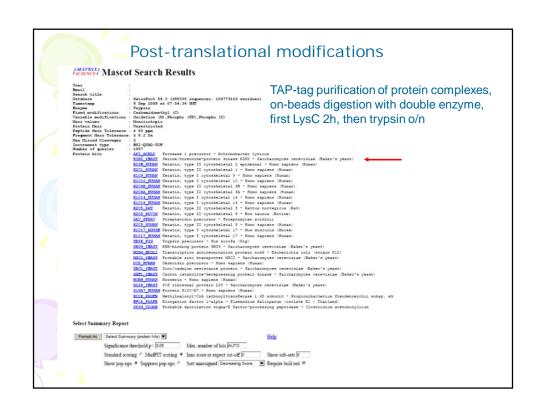
Elias, J. E., et al, Nature Methods 2 667-675 (2005).

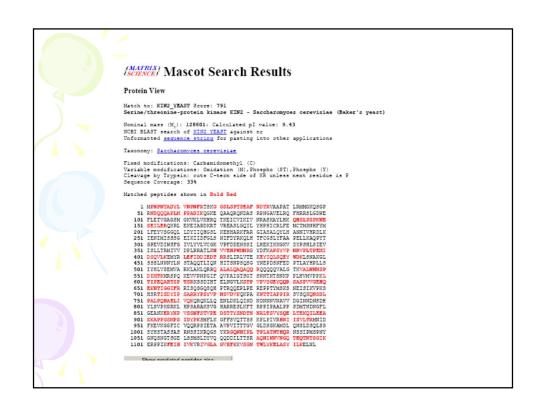
Now you're protein is identified What else can we find in the data?

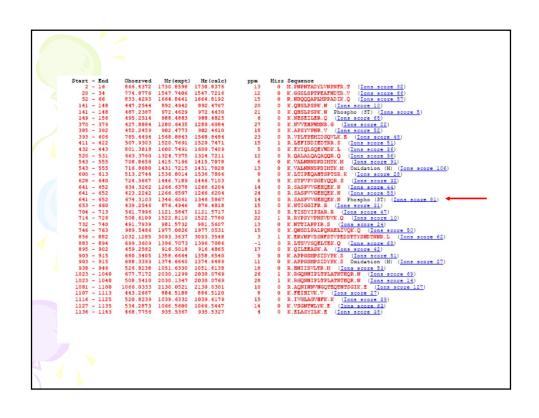
Protein fragmentation
-protein separation by SDS-PAGE/2-DE
followed by in-gel digestion and MS analysis

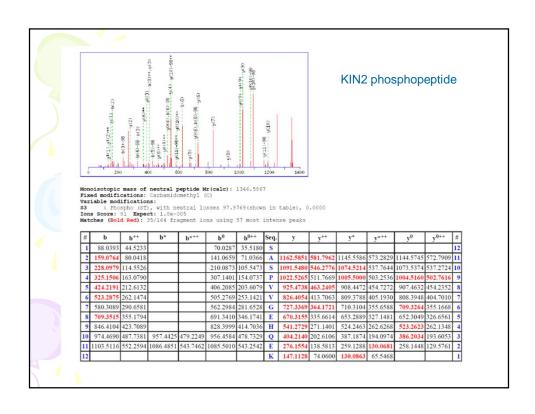












De novo sequencing

De novo = peptide sequencing performed without prior knowledge of the amino acid sequence

- -if enough material is available classical Edman degradation is still a vey good method
- -partial peptide sequencing is possible based on MS/MS data

