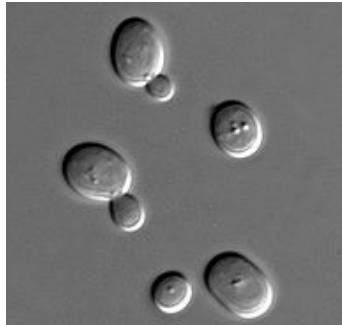


Phosphoproteomics

PhD Henri Blomster

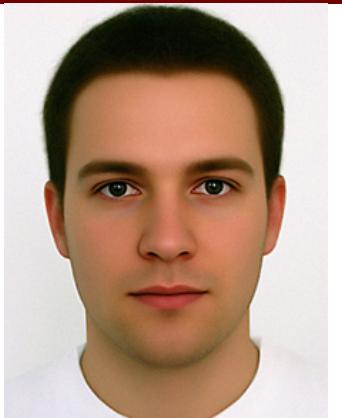
Human Genome Project



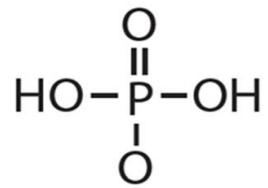
Budding yeast 6,000 genes



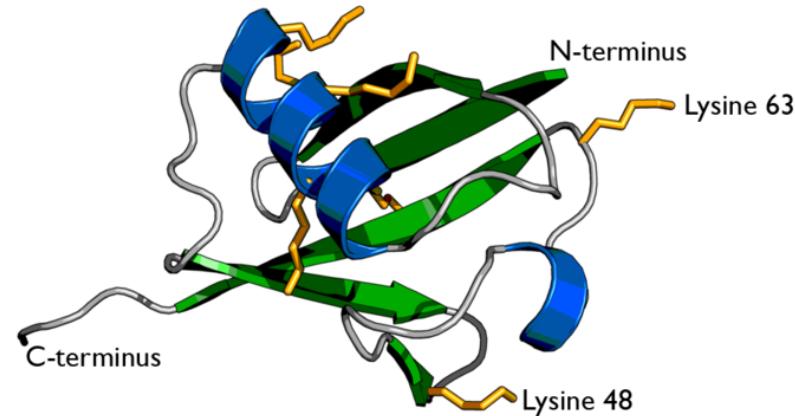
Nematode 15,000 genes



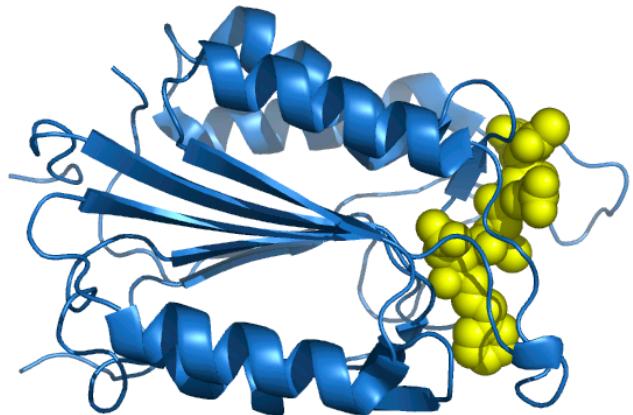
Human 25,000 genes



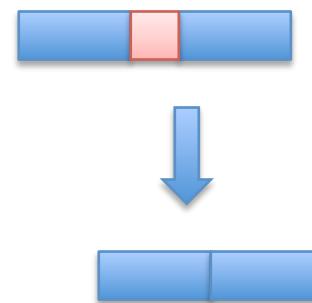
Attachment of chemical groups



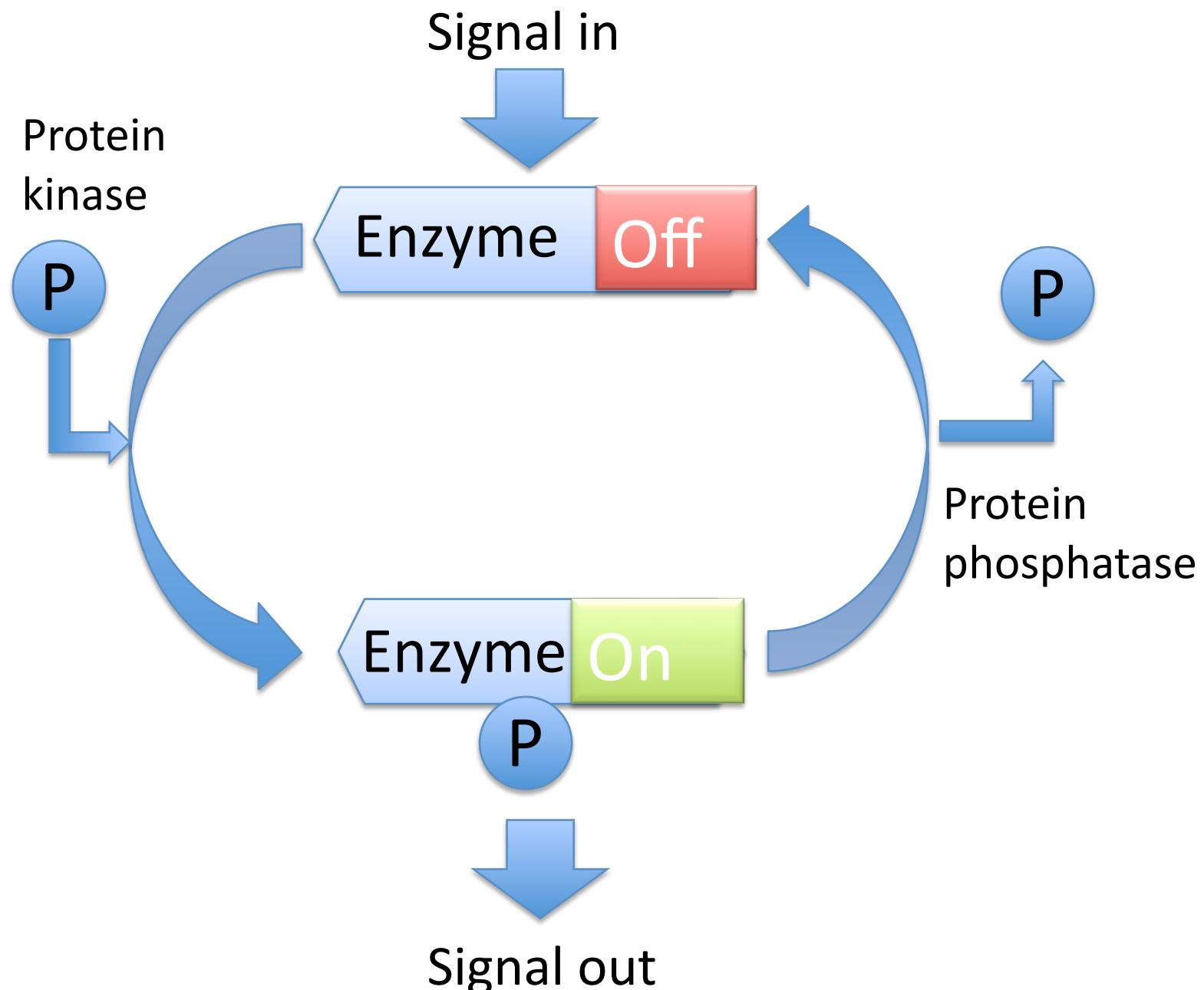
Covalent attachment of polypeptides



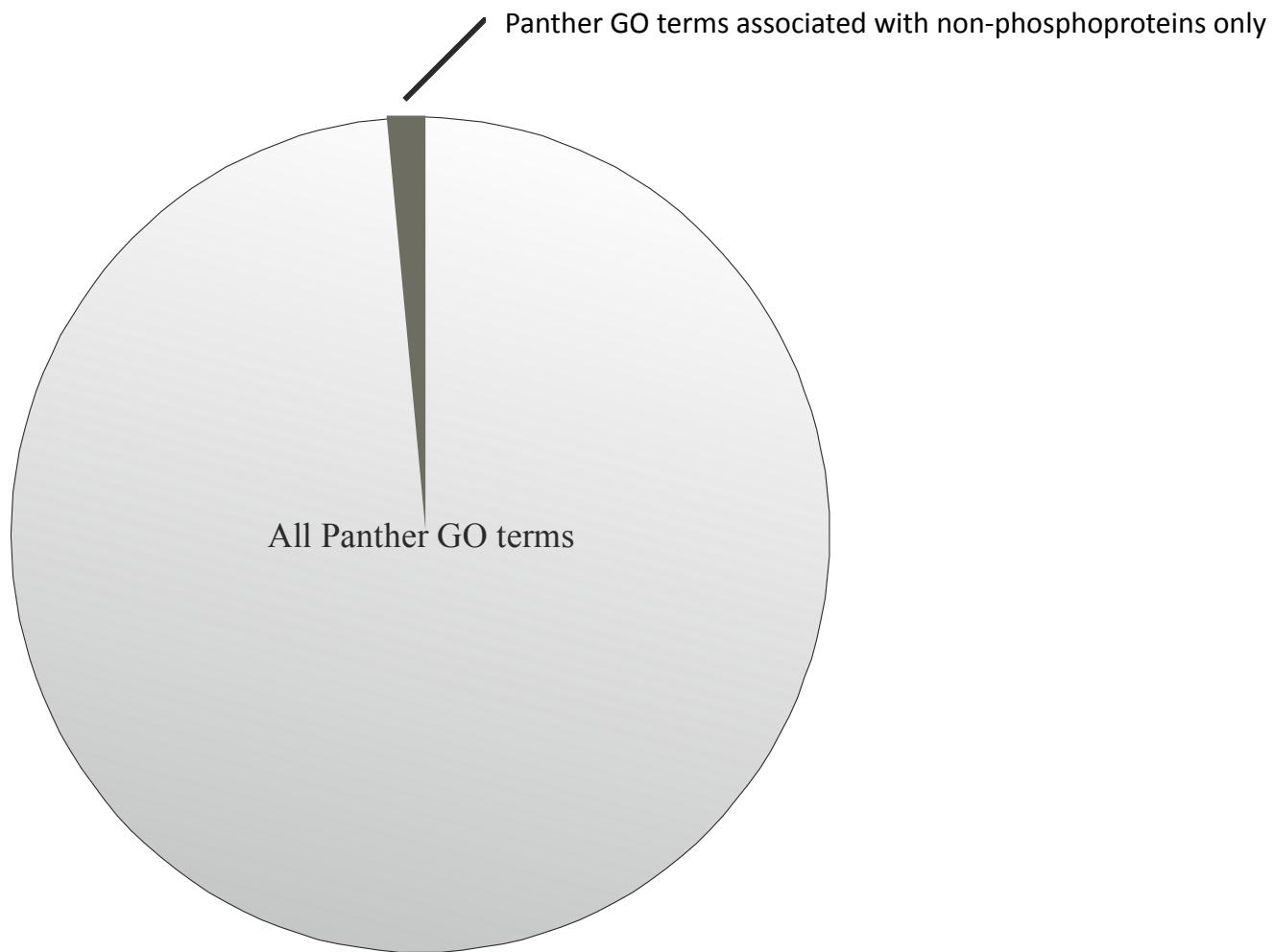
Protein cleavage

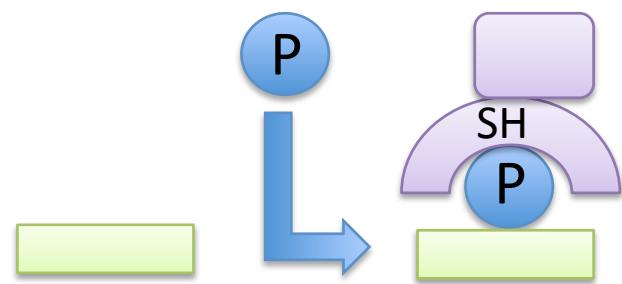


Protein splicing

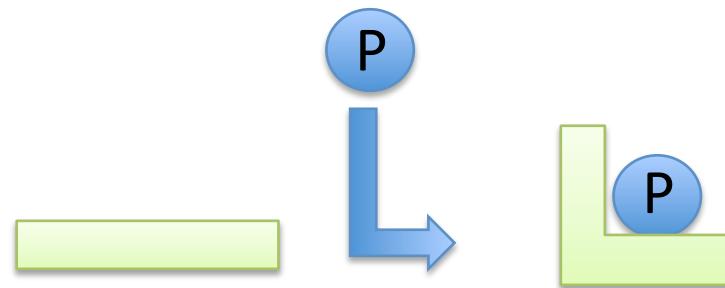


Why to map PTM sites: a systems biology view

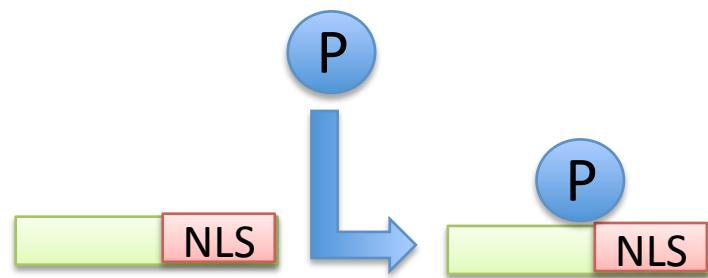




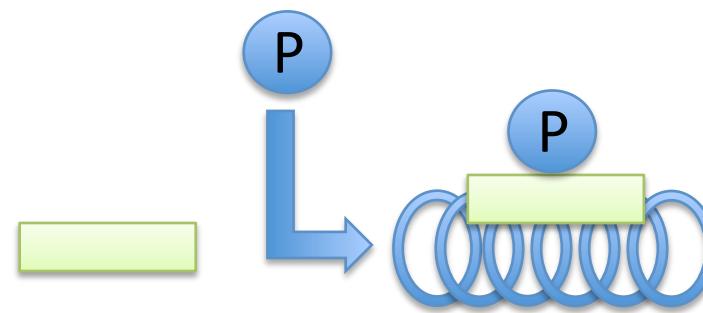
Recruitment of effector proteins



Conformational change

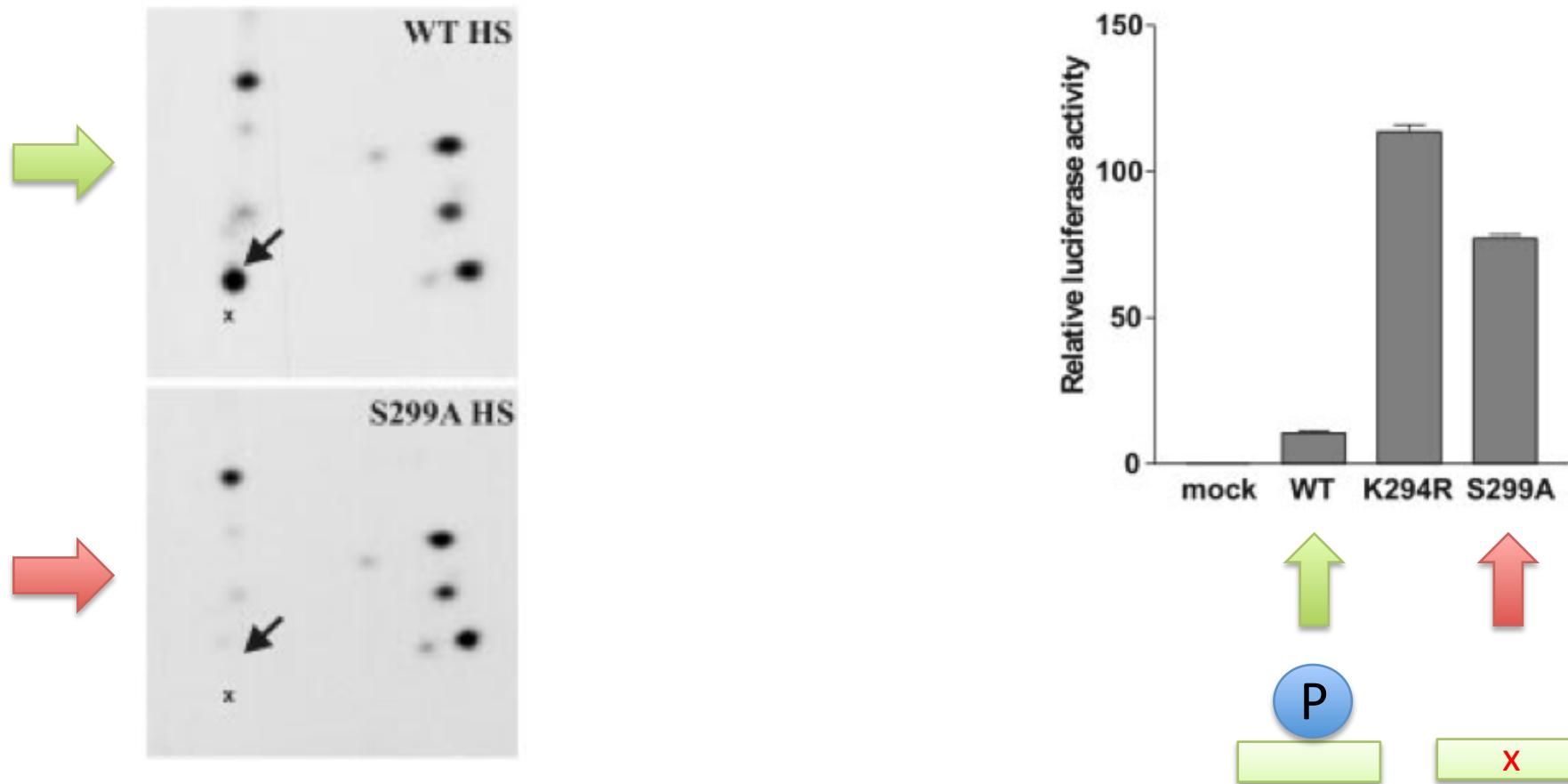


Cellular localisation



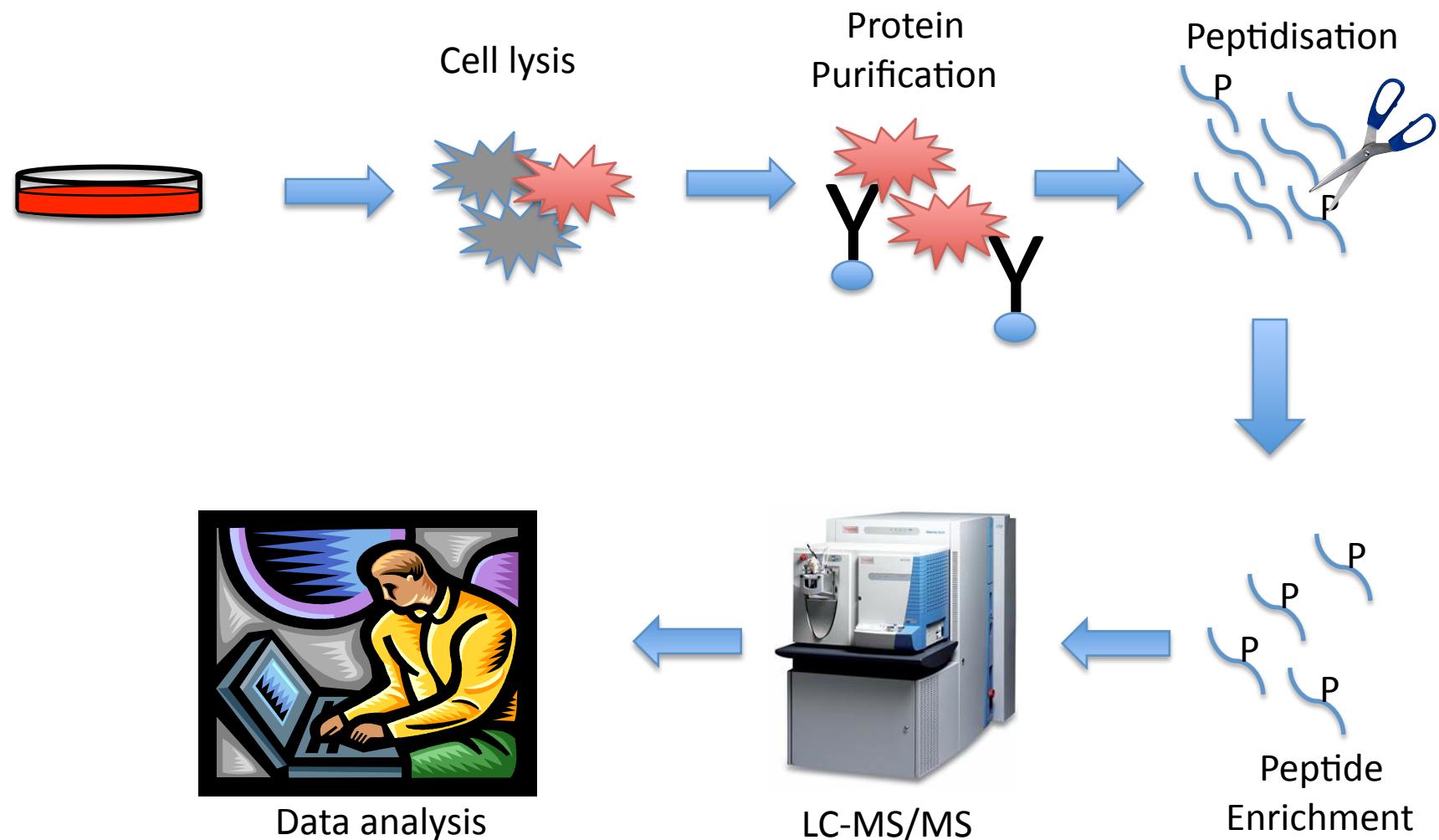
DNA binding

Why to map PTM sites: a single protein's view, HSF4b



Hietakangas et al. PNAS, 2006

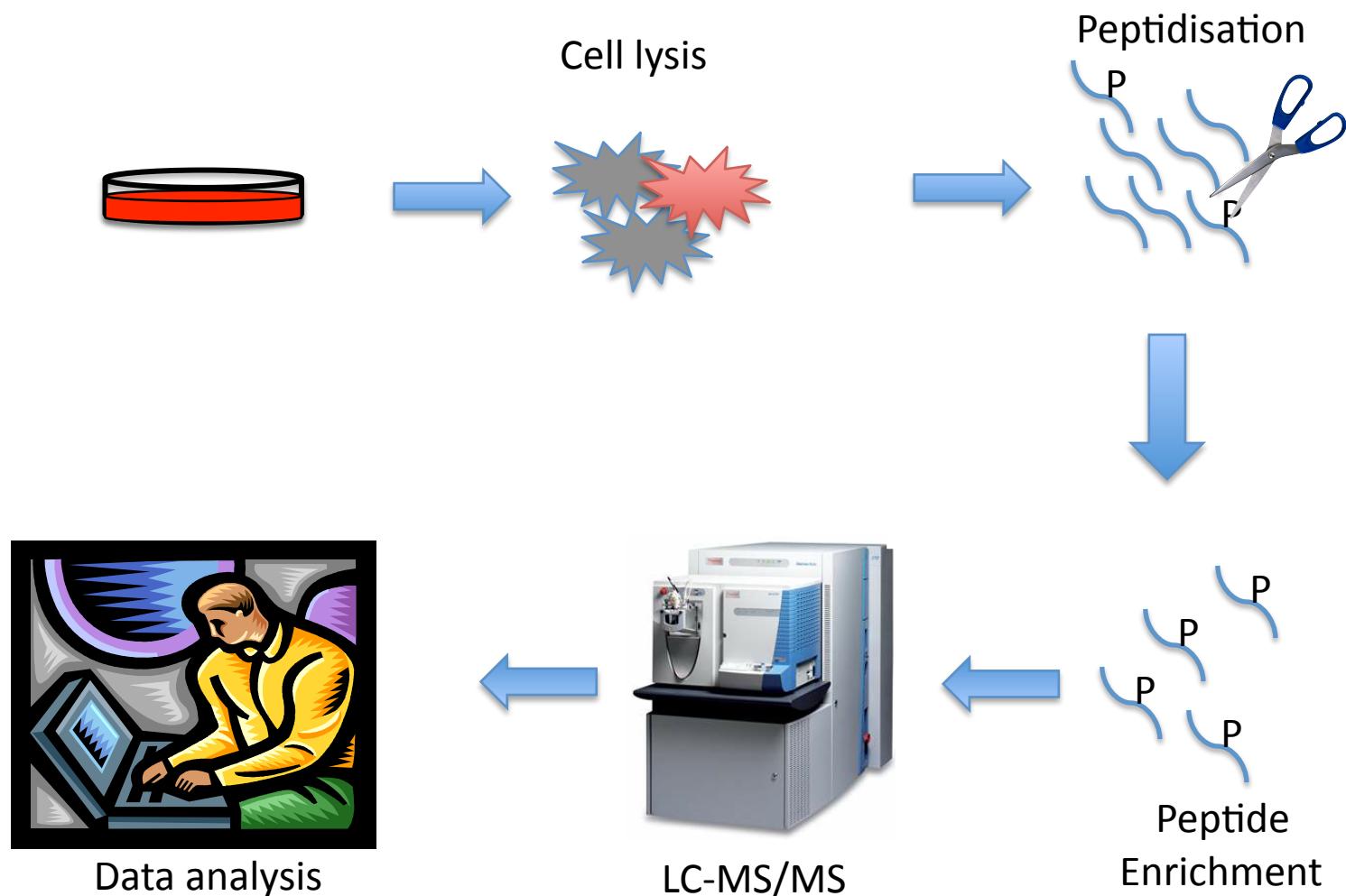
A targeted phosphoproteomics approach



A targeted phosphoproteomics approach

- Advantages
 - Sensitive approach especially for proteins that are expressed at low level like transcription factors
- Disadvantages
 - Low throughput
 - Detect only things that are looked at

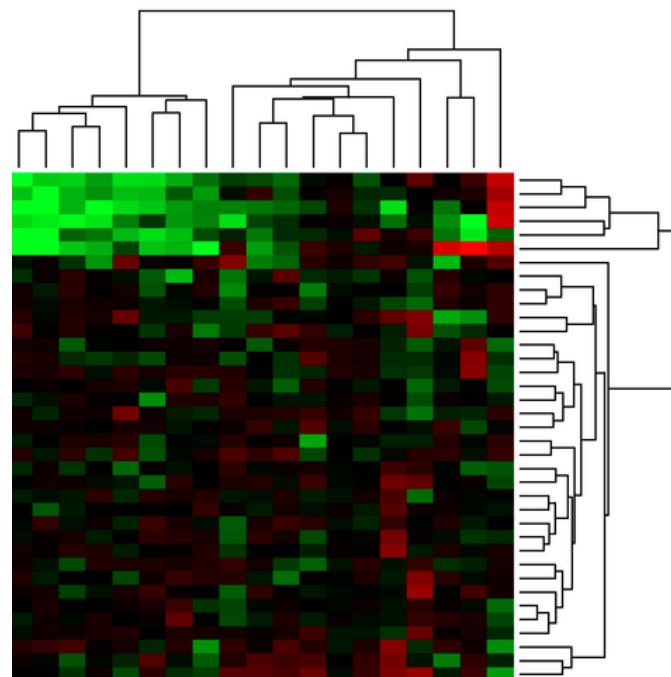
Global phosphoproteomics approach



A global approach

- Advantages
 - high number of phosphorylation sites identified
- Disadvantages
 - low-abundant proteins not detected

The phosphoproteome is dynamic



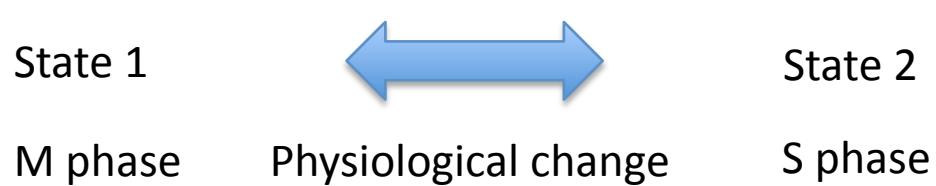
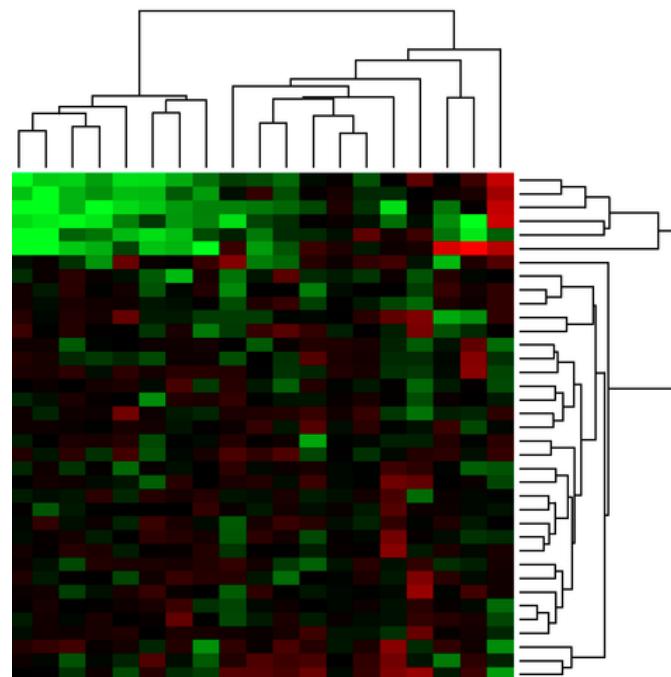
State 1



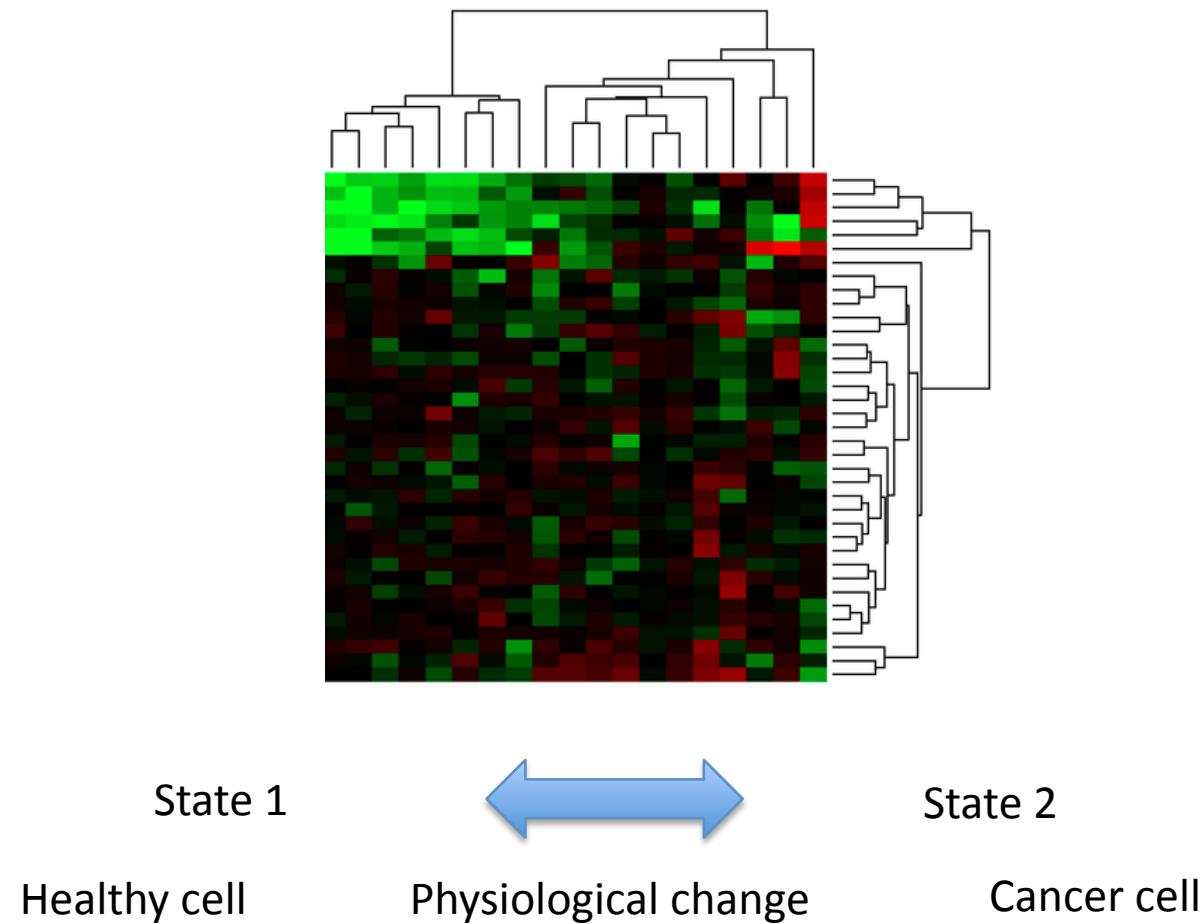
State 2

Physiological change

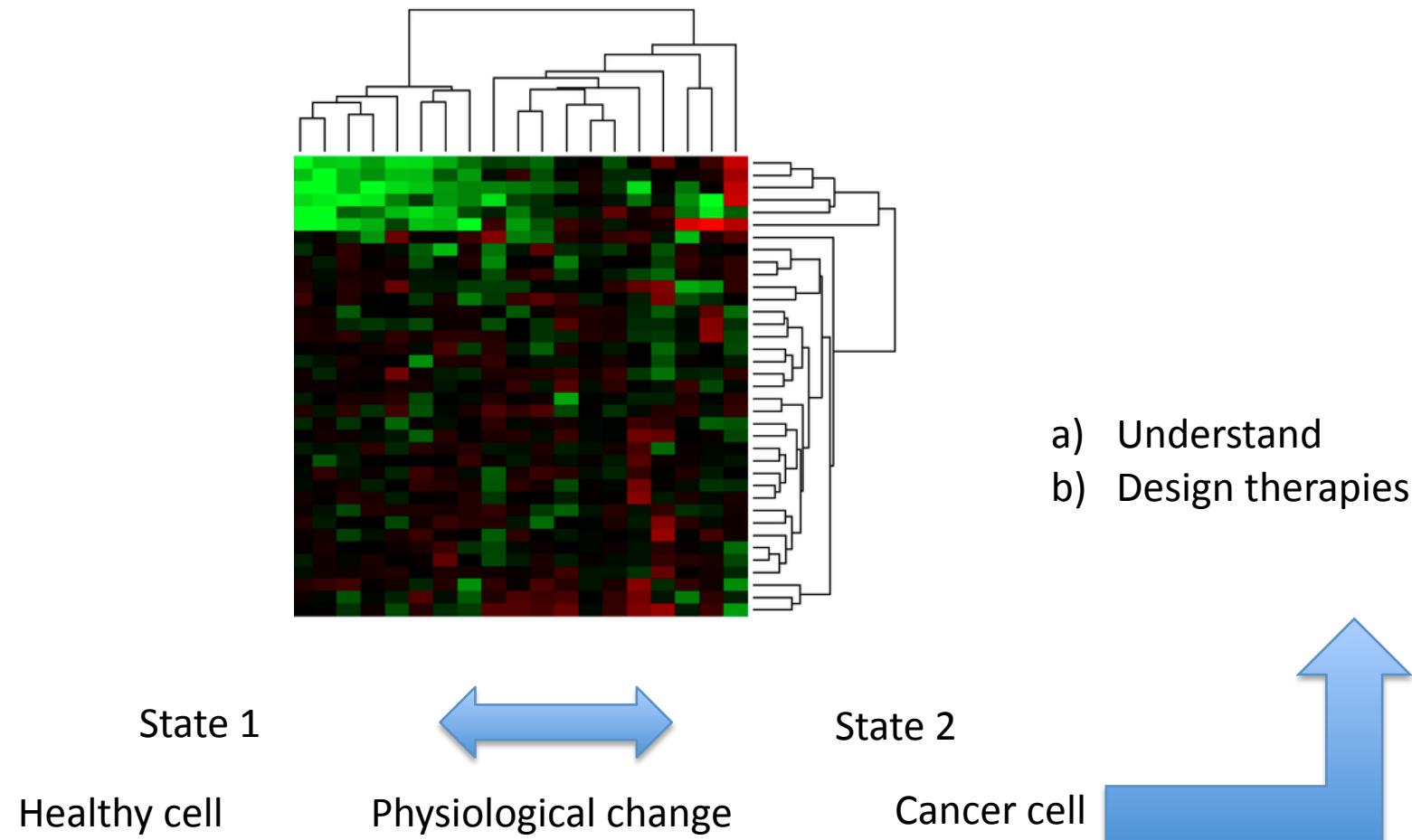
The phosphoproteome is dynamic



The phosphoproteome is dynamic



The phosphoproteome is dynamic

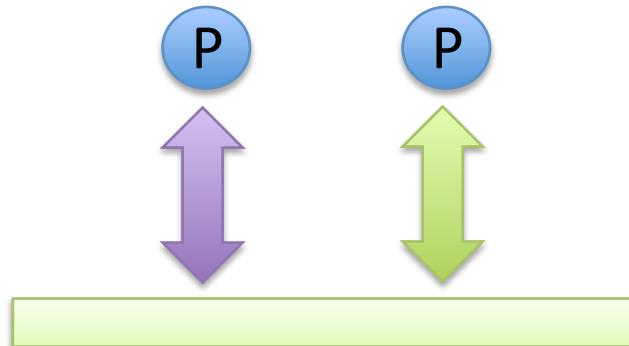


Why to quantitate, a single proteins view

- Biological function is difficult to find
 - Many phosphorylation sites are background
 - Many biological functions
- Quantitative proteomics is a good starting point

Why to quantitate, a global view

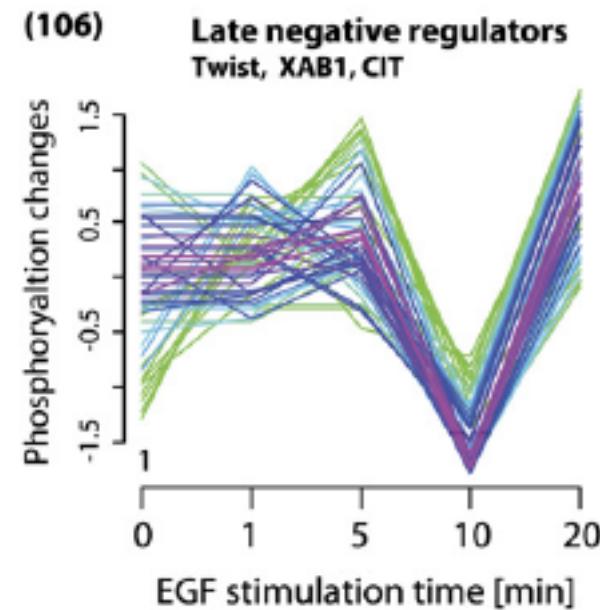
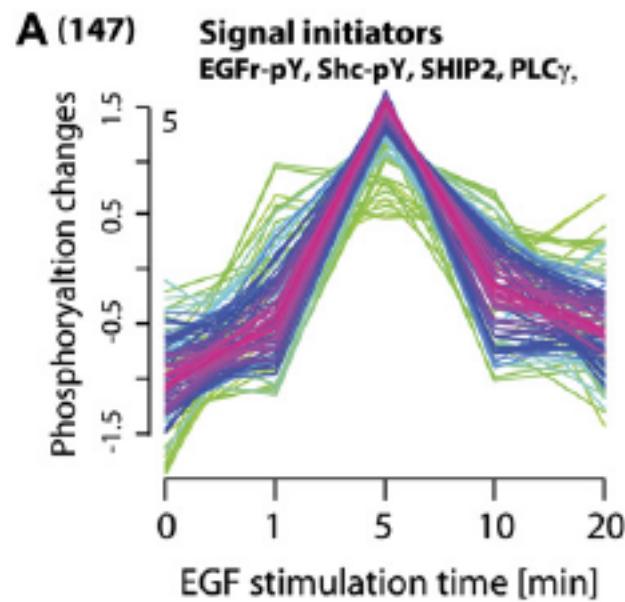
Global quantitation of phosphorylation sites after growth factor stimulation



Olsen et al. *Cell* 2006

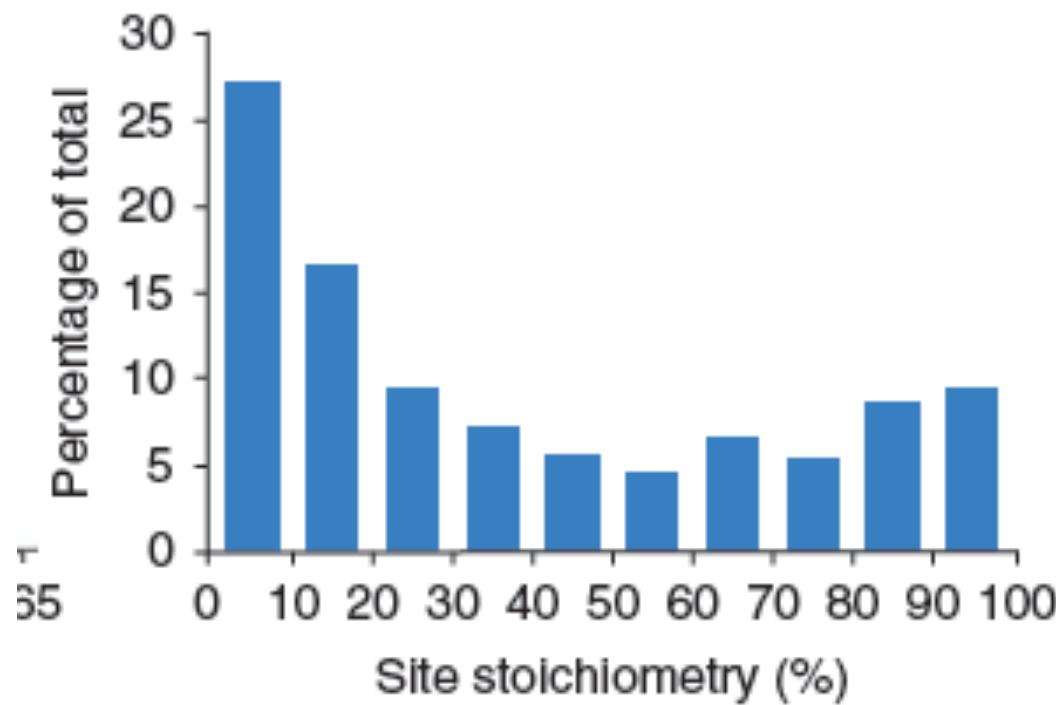
Why to quantitate, a global view

Global quantitation of phosphorylation sites after growth factor stimulation



Classification by temporal profiles

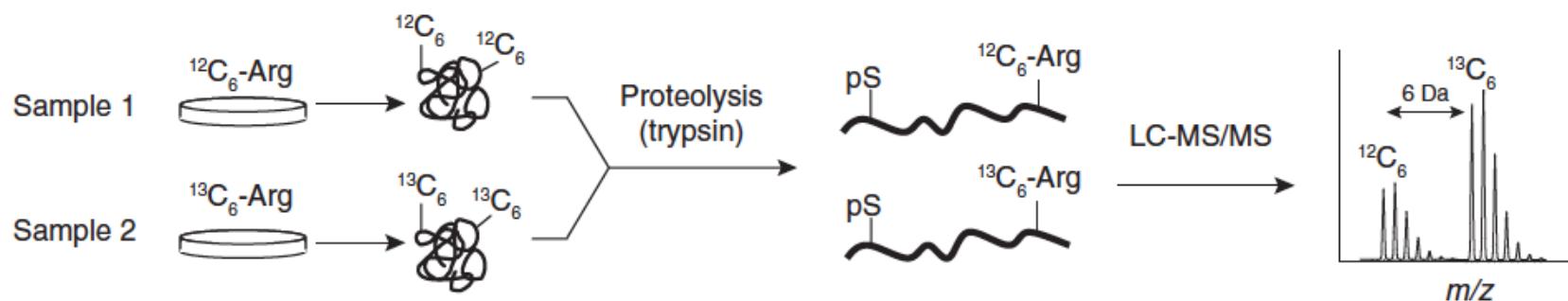
Olsen et al. *Cell* 2006

C

Site stoichiometry distribution for 5,033 events
from wild-type yeast undergoing exponential
growth

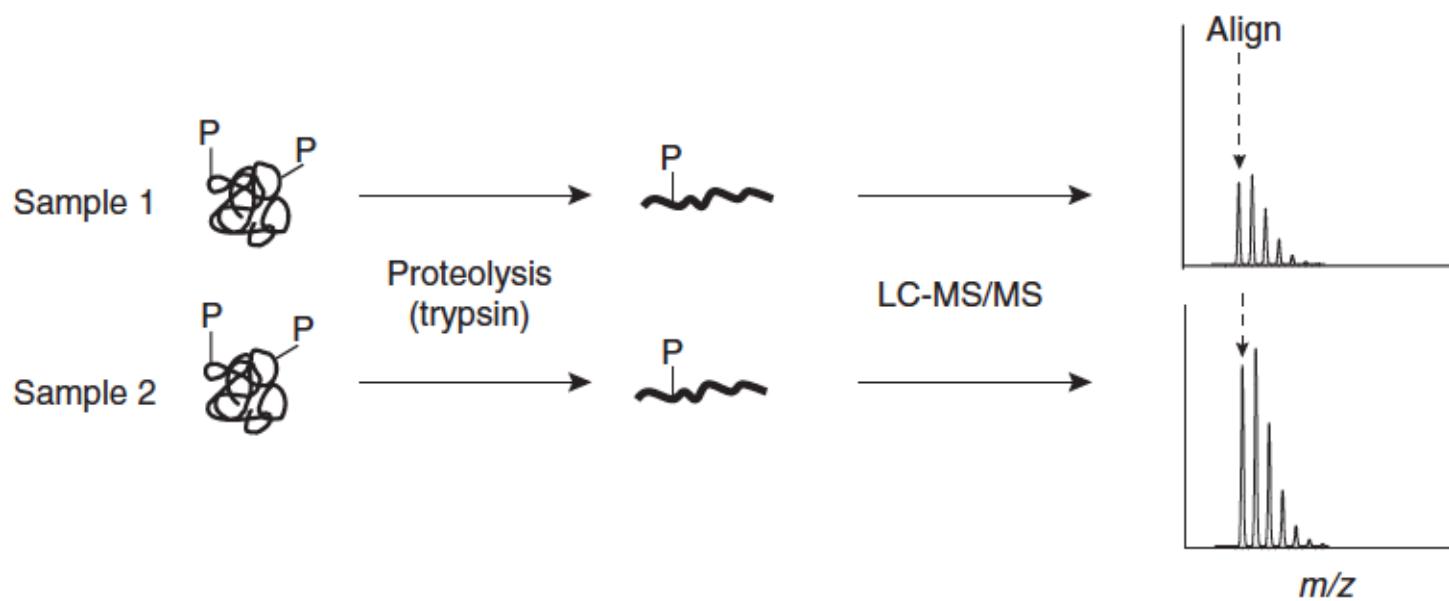
a

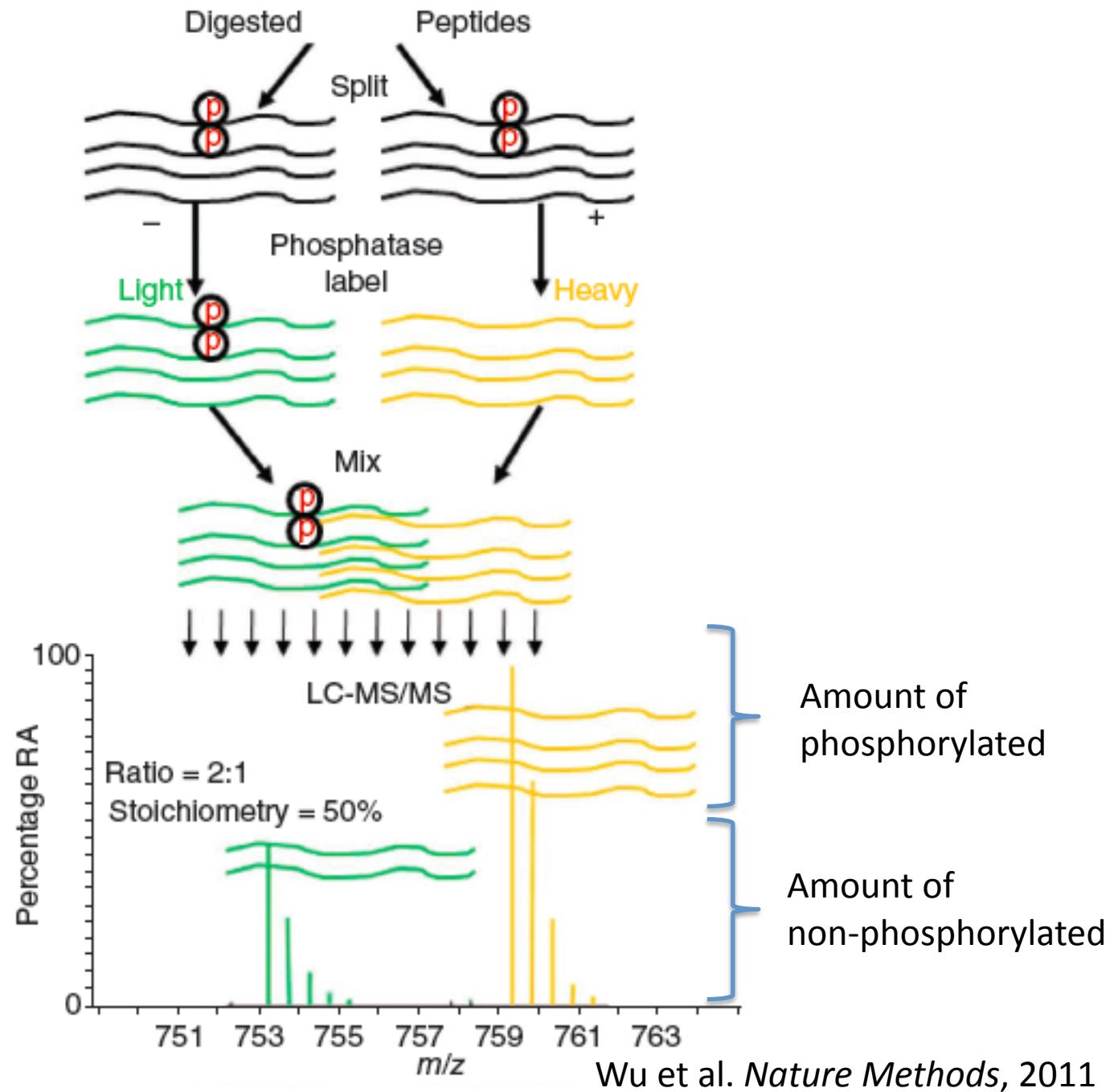
Metabolic labeling



b

Peak alignment
and intensity
measurement



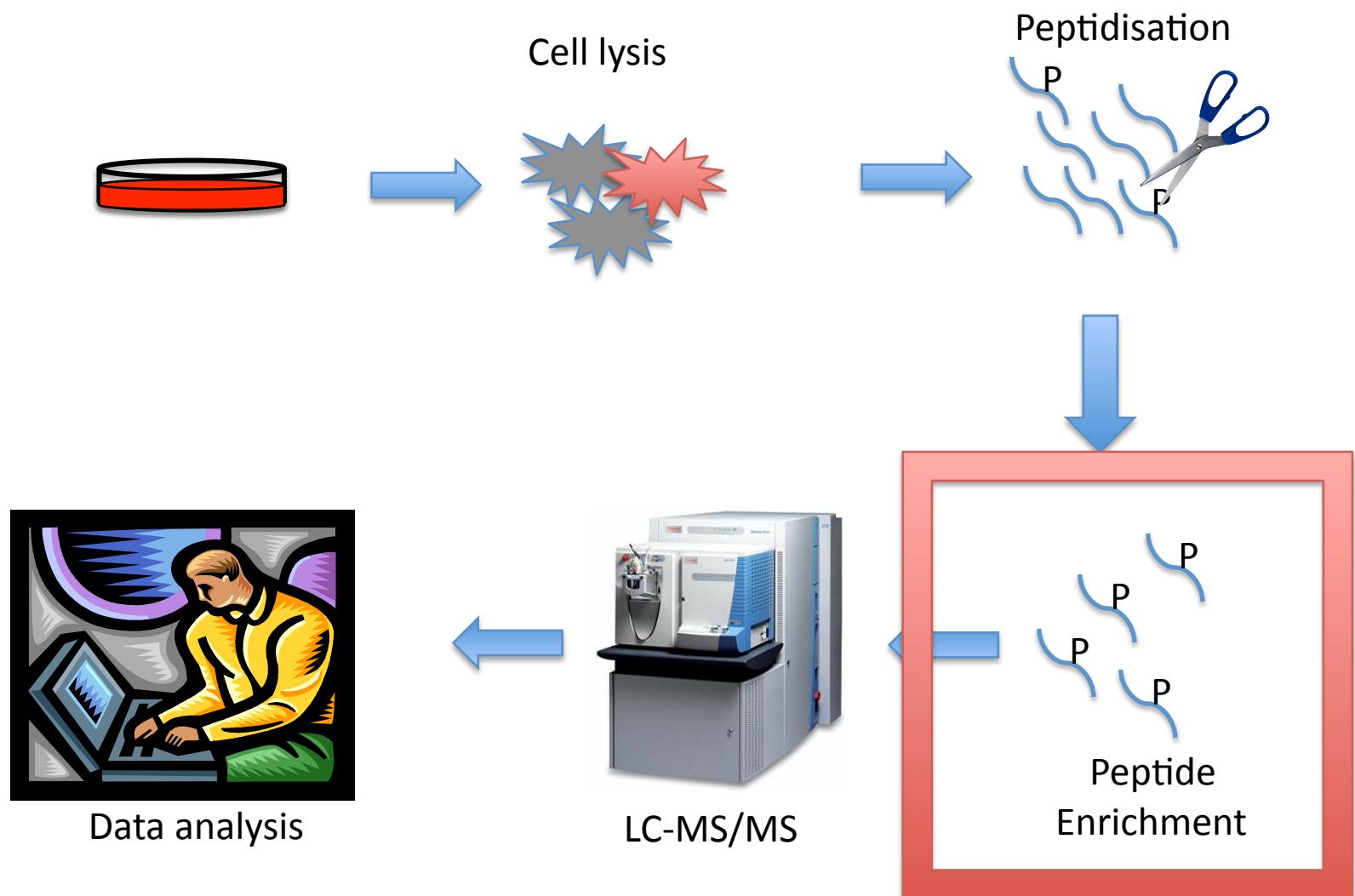


Summary of Part I

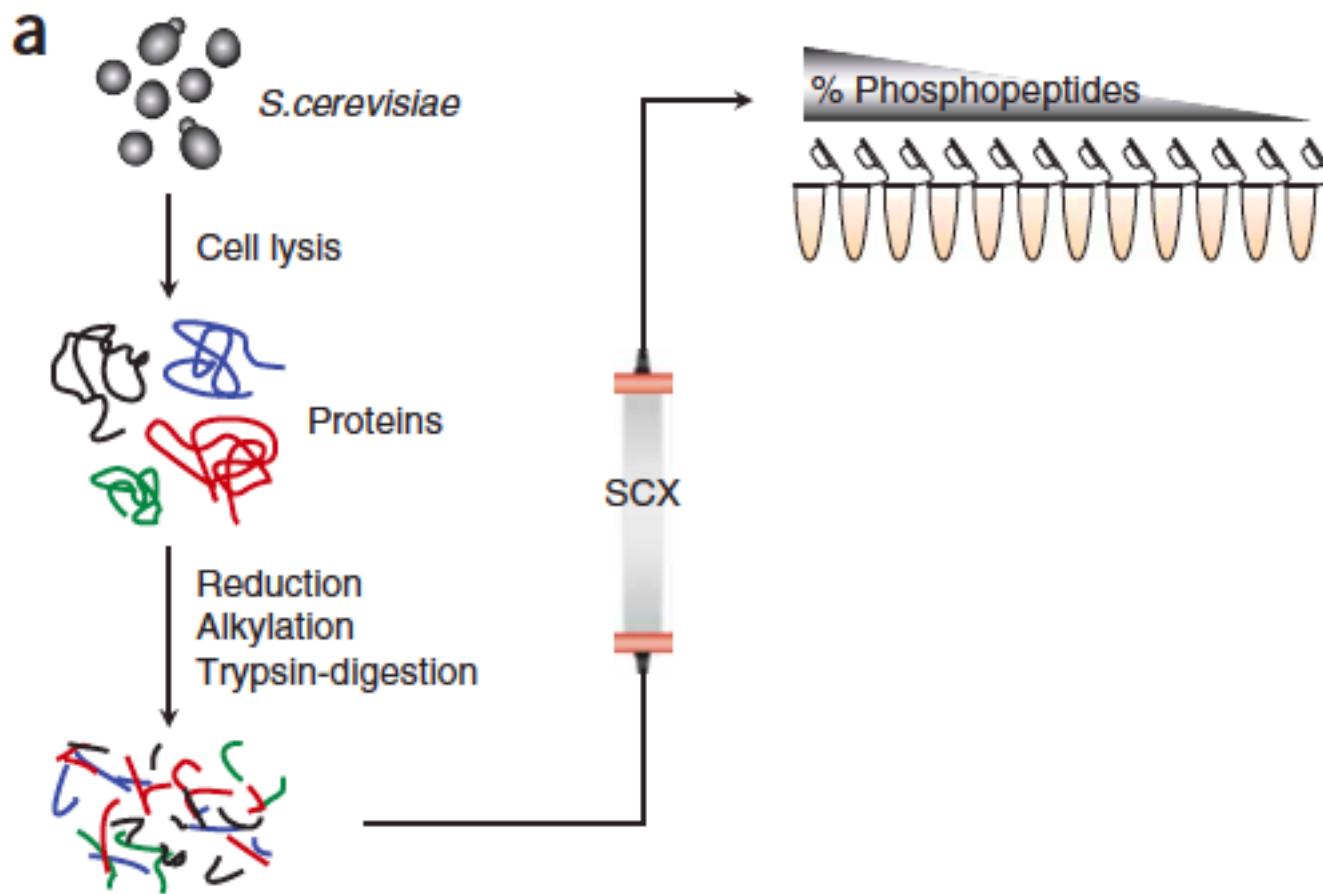
- Importance of PTM's
- Why to map PTM sites
 - targeted approach
 - global approach
- How to map PTM sites
 - targeted approach
 - global approach

How to map phosphorylation sites with mass spectrometry

Global phosphoproteomics approach

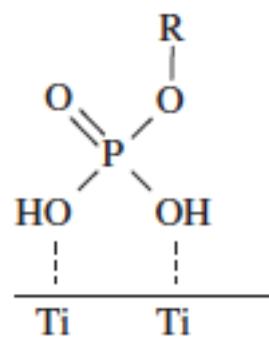
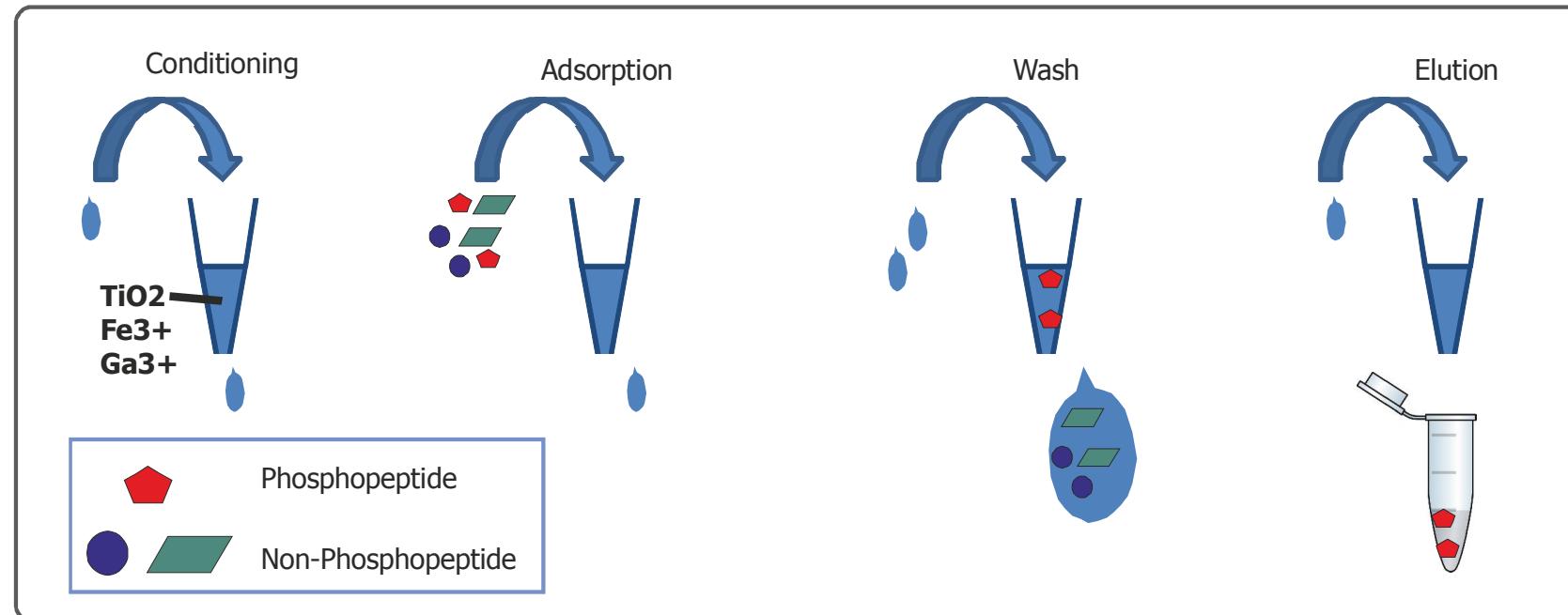


Strong cation exchange, SCX separates peptides by charge. Phosphate groups add negative charge and phosphopeptides are expected to elute earlier



IMAC and TiO₂ enrichment

Immobilised metal ion chromatography (IMAC) makes use of matrix-bound metals to affinity purify phosphopeptides. (positive metal, negative phosphate)



Titanium dioxide affinity purification makes use of matrix-bound TiO_2 to affinity purify phosphopeptides.

Engholm-Keller et al. *Journal of Proteomics*, 2011

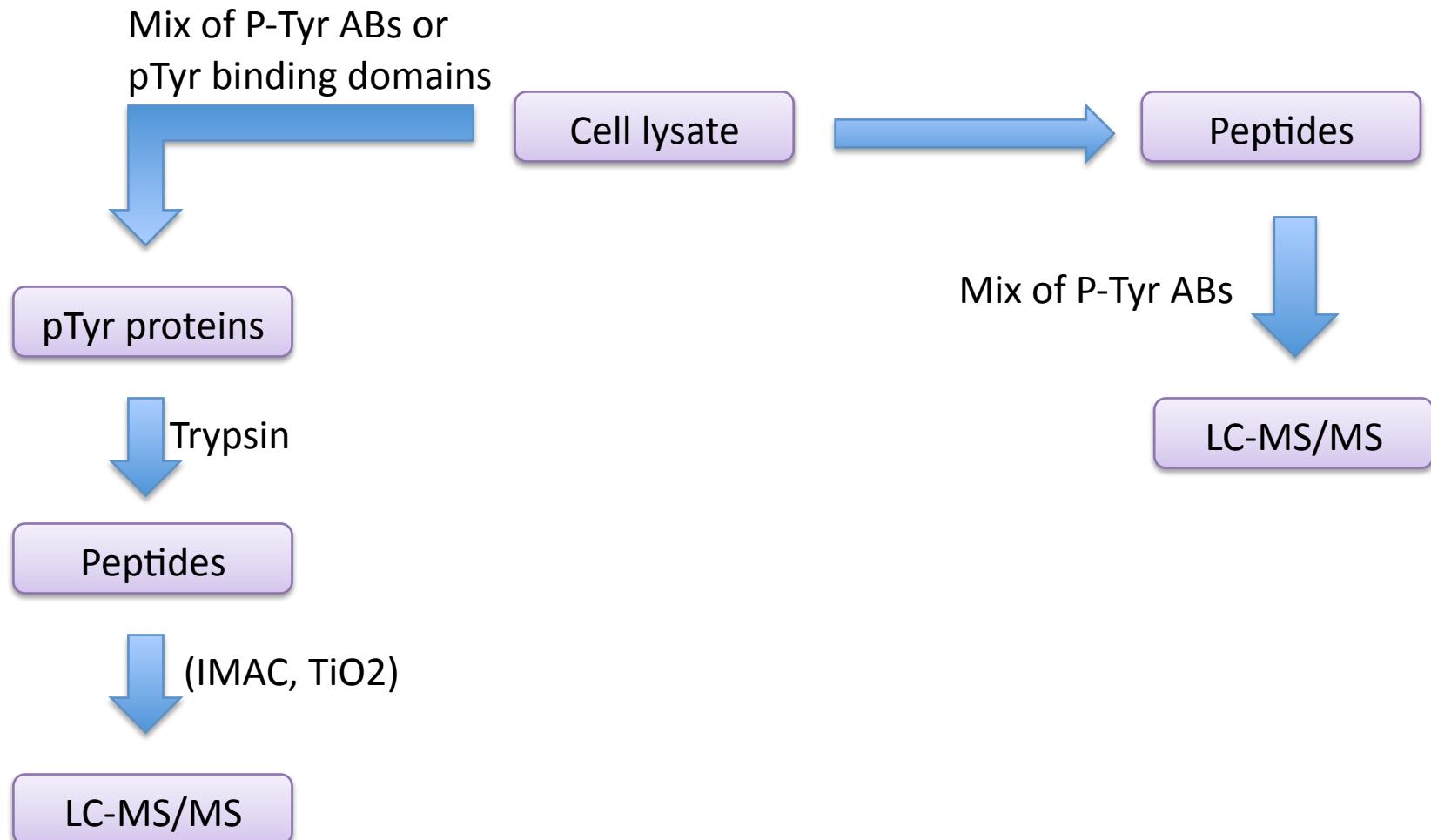
Phosphotyrosine signalling

Distribution of phosphorylation sites by amino acid

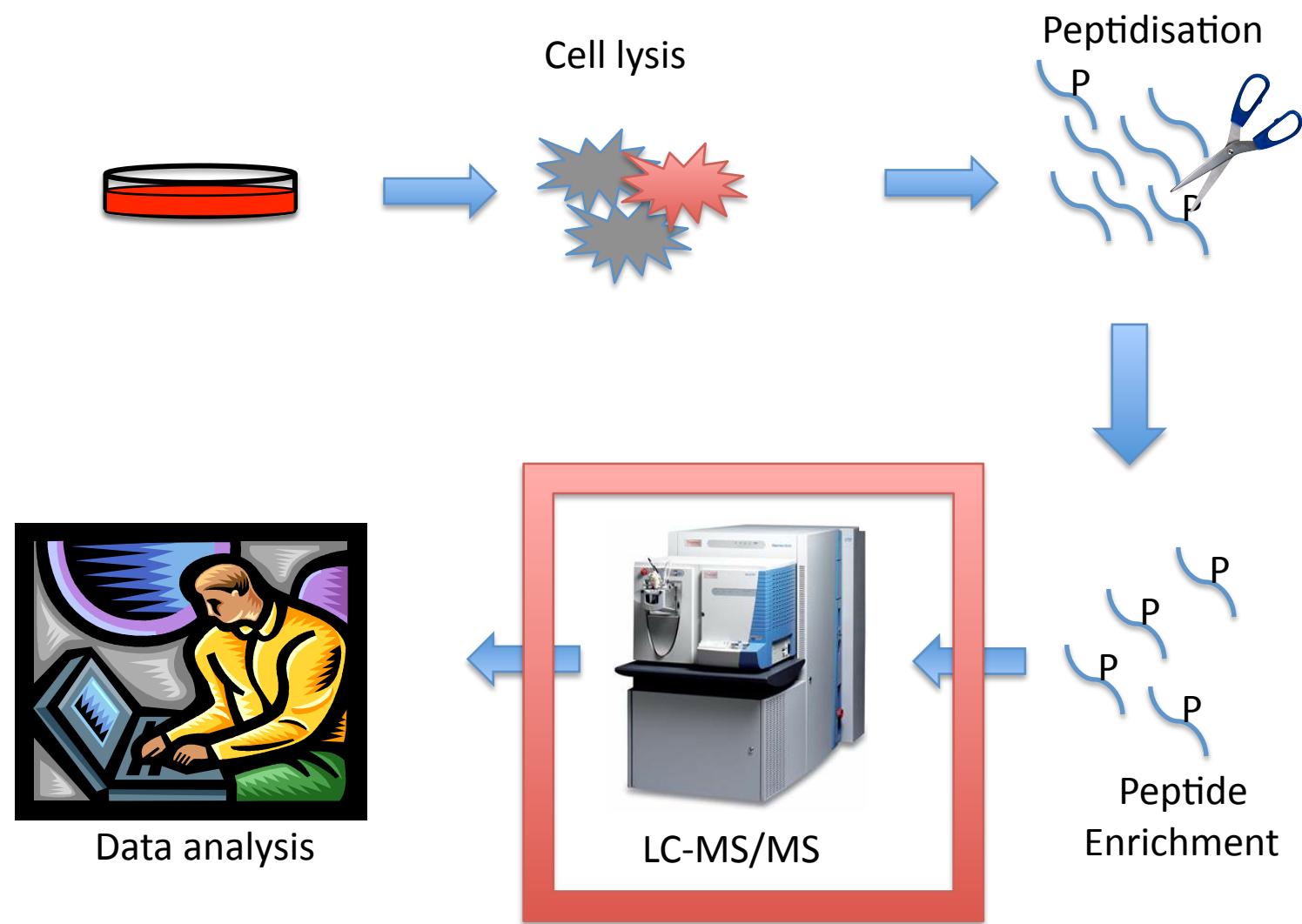
Site	Class I	Percent	EGF-regulated	Percent
pSer	4901	86.4%	724	82.0%
pThr	670	11.8%	106	12.0%
pTyr	103	1.8%	53	6.0%

Olsen et al. *Cell* 2006

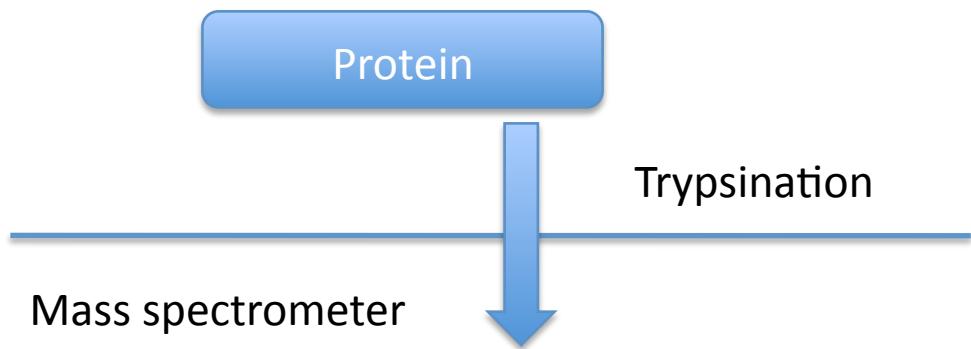
Phosphotyrosine signalling



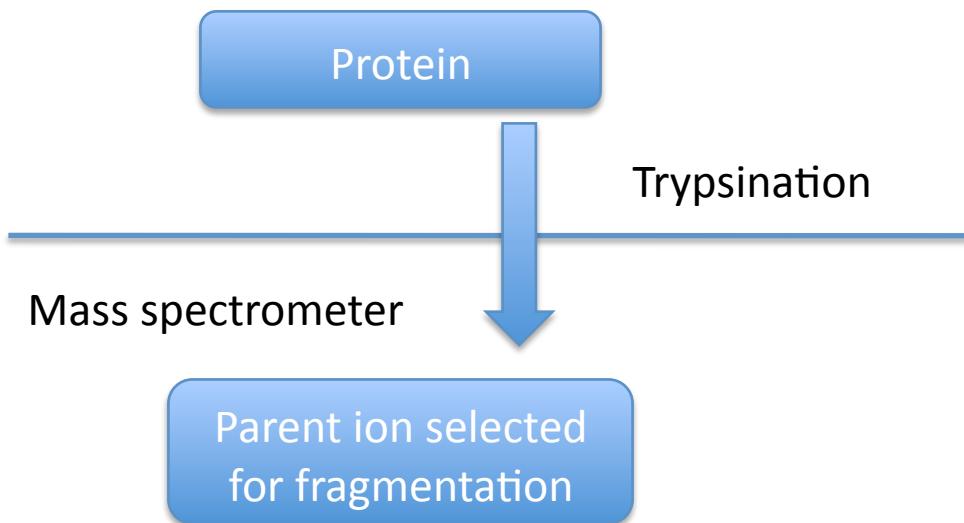
Global phosphoproteomics approach



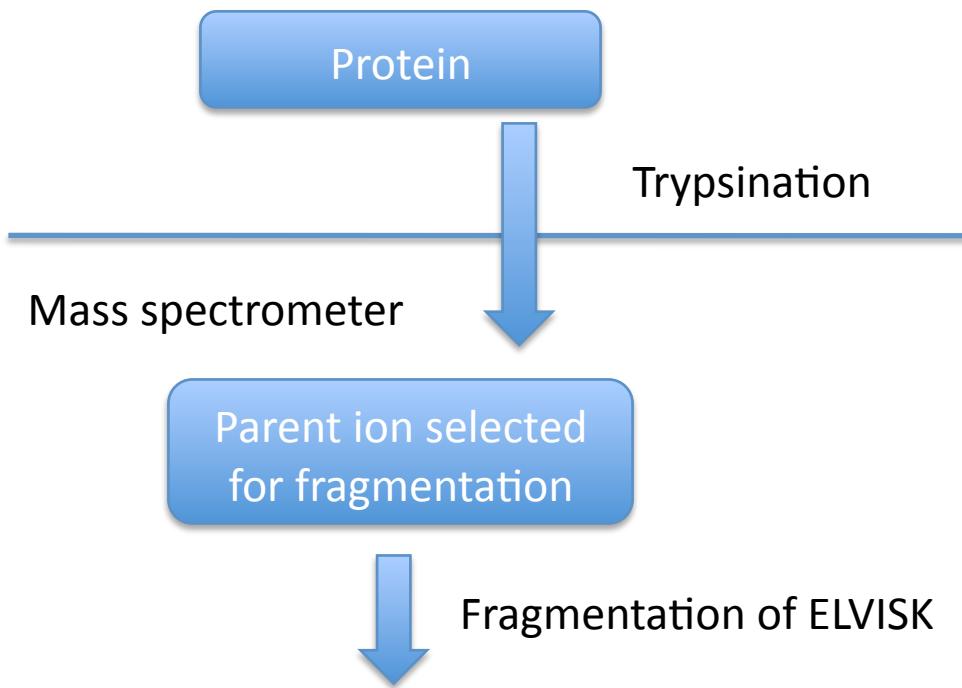
Wet lab



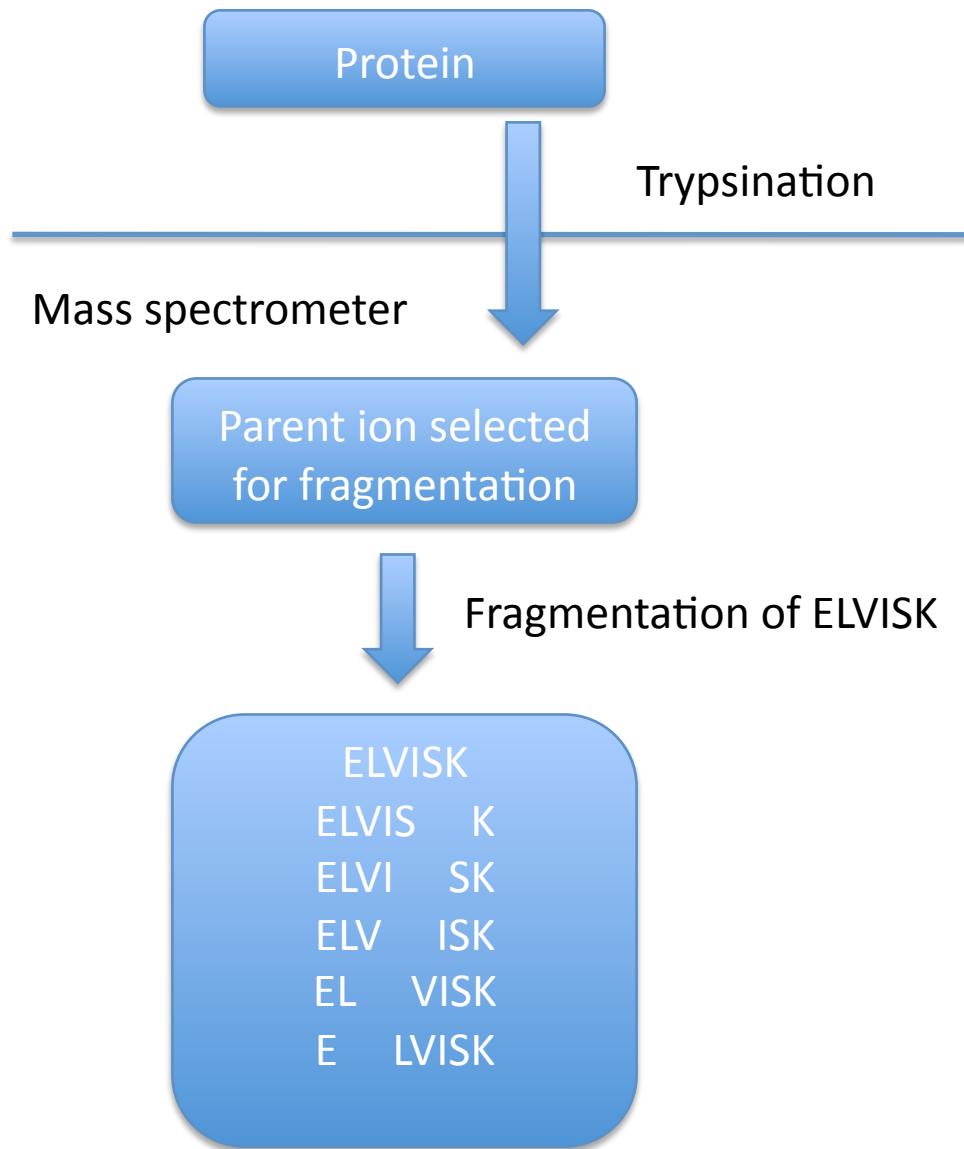
Wet lab



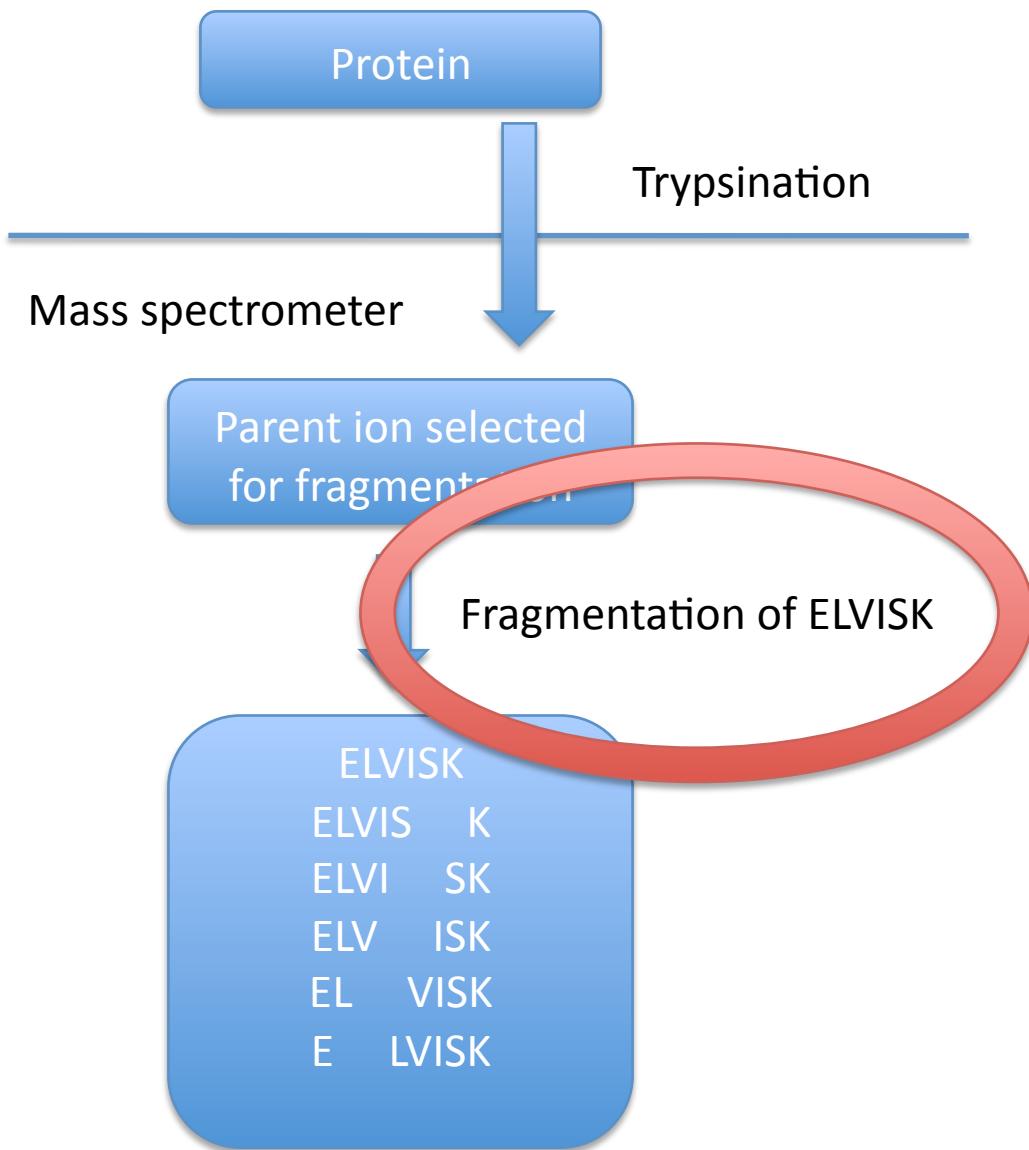
Wet lab



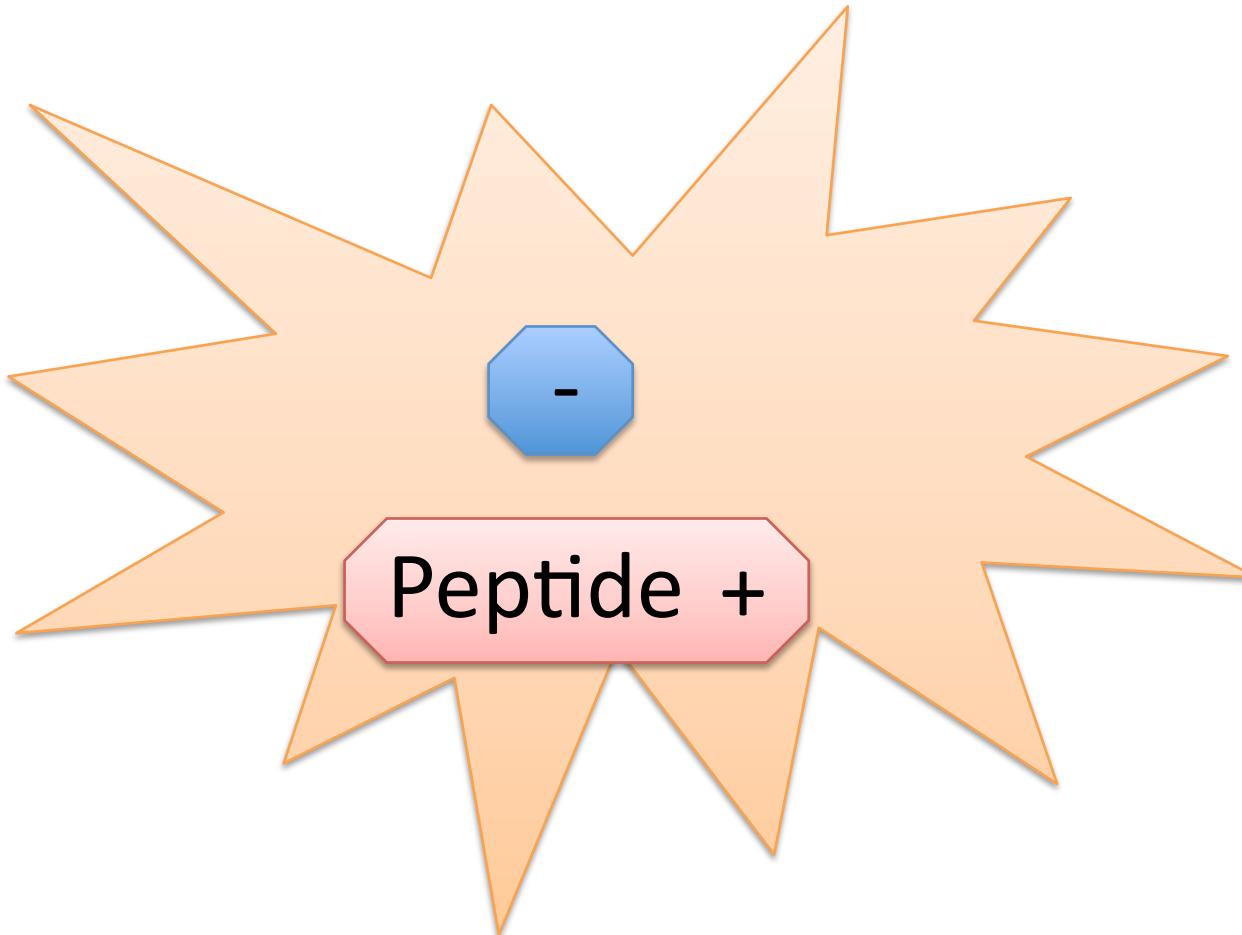
Wet lab



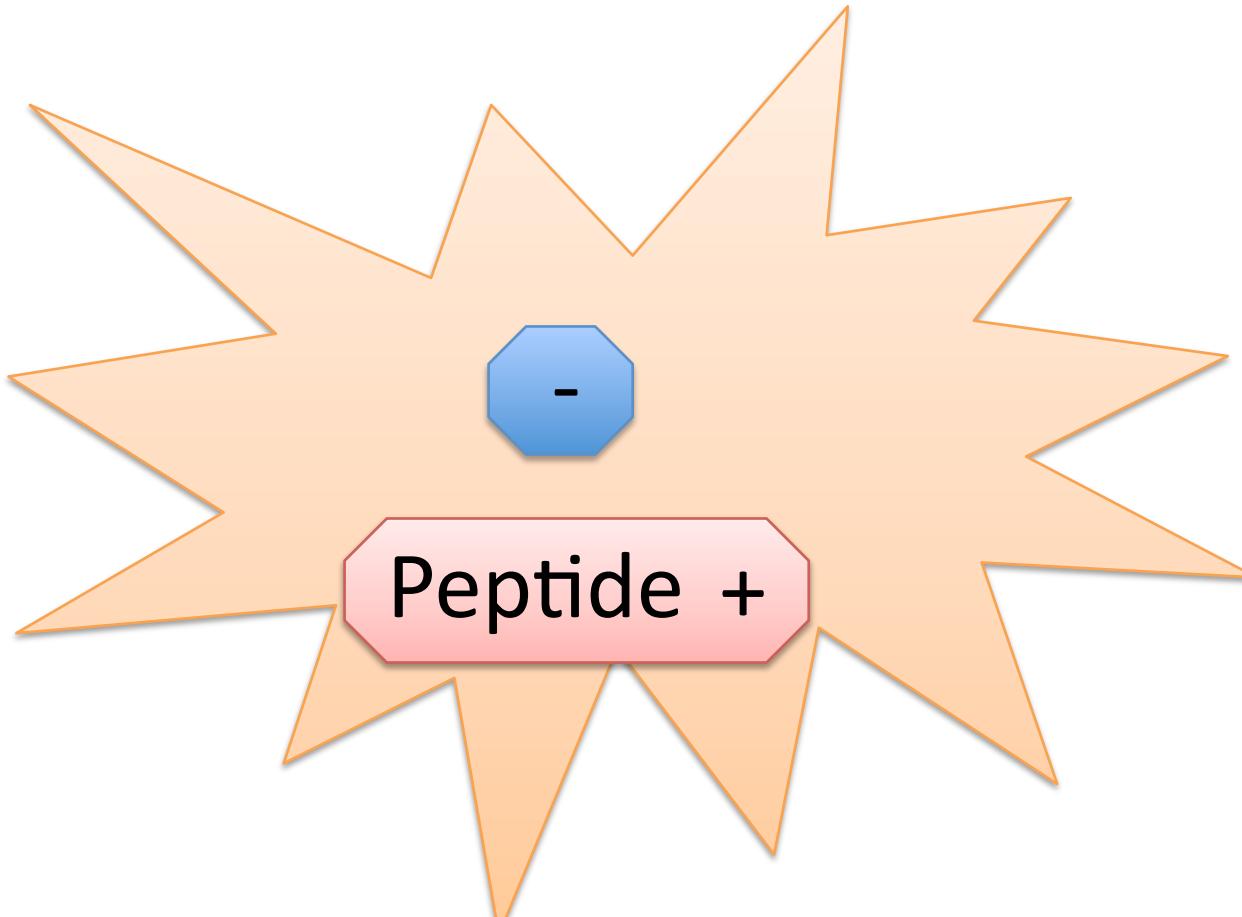
Wet lab



Electron transfer dissociation ETD



Electron transfer dissociation ETD

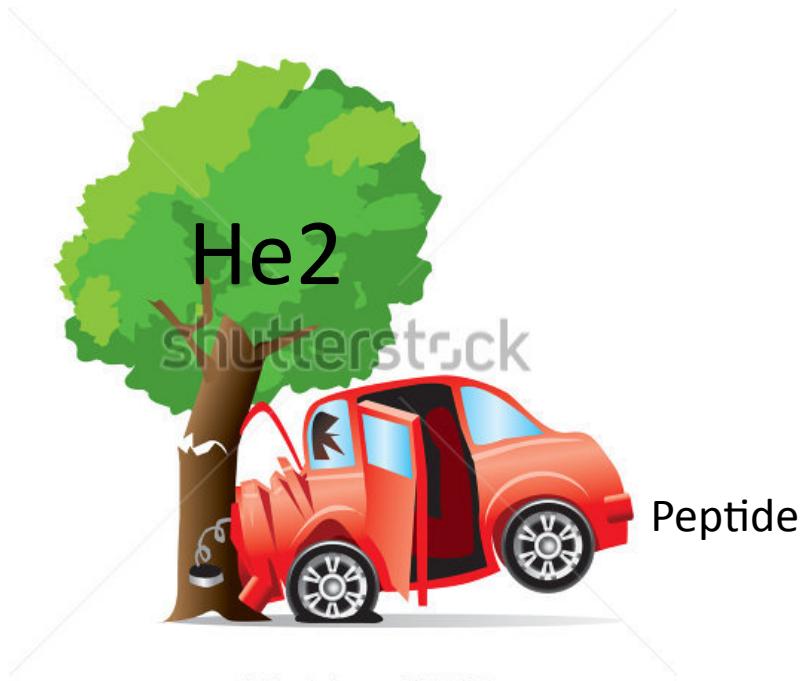


ETD, no neutral loss of H₃PO₄

Collision induced dissociation CID

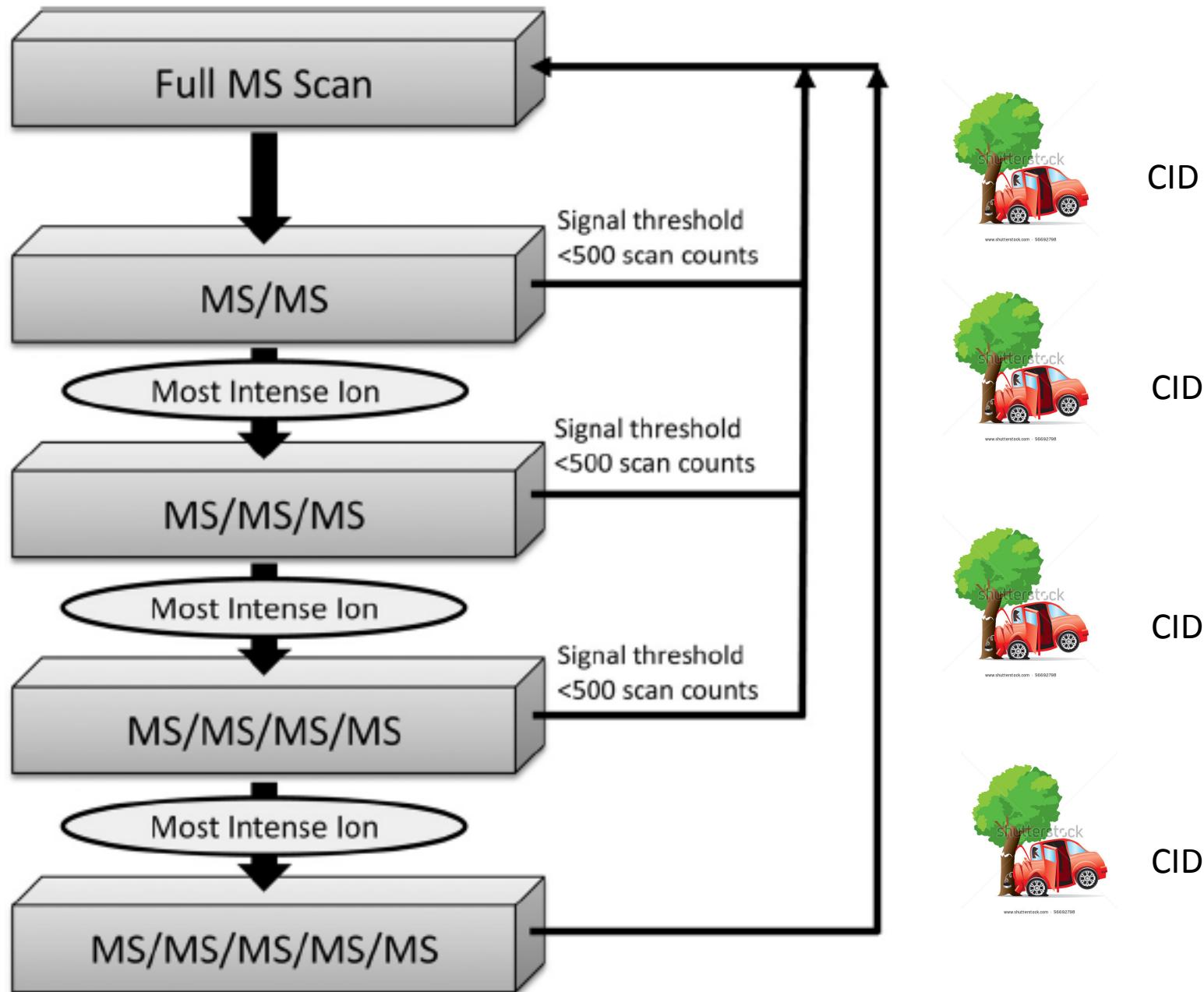


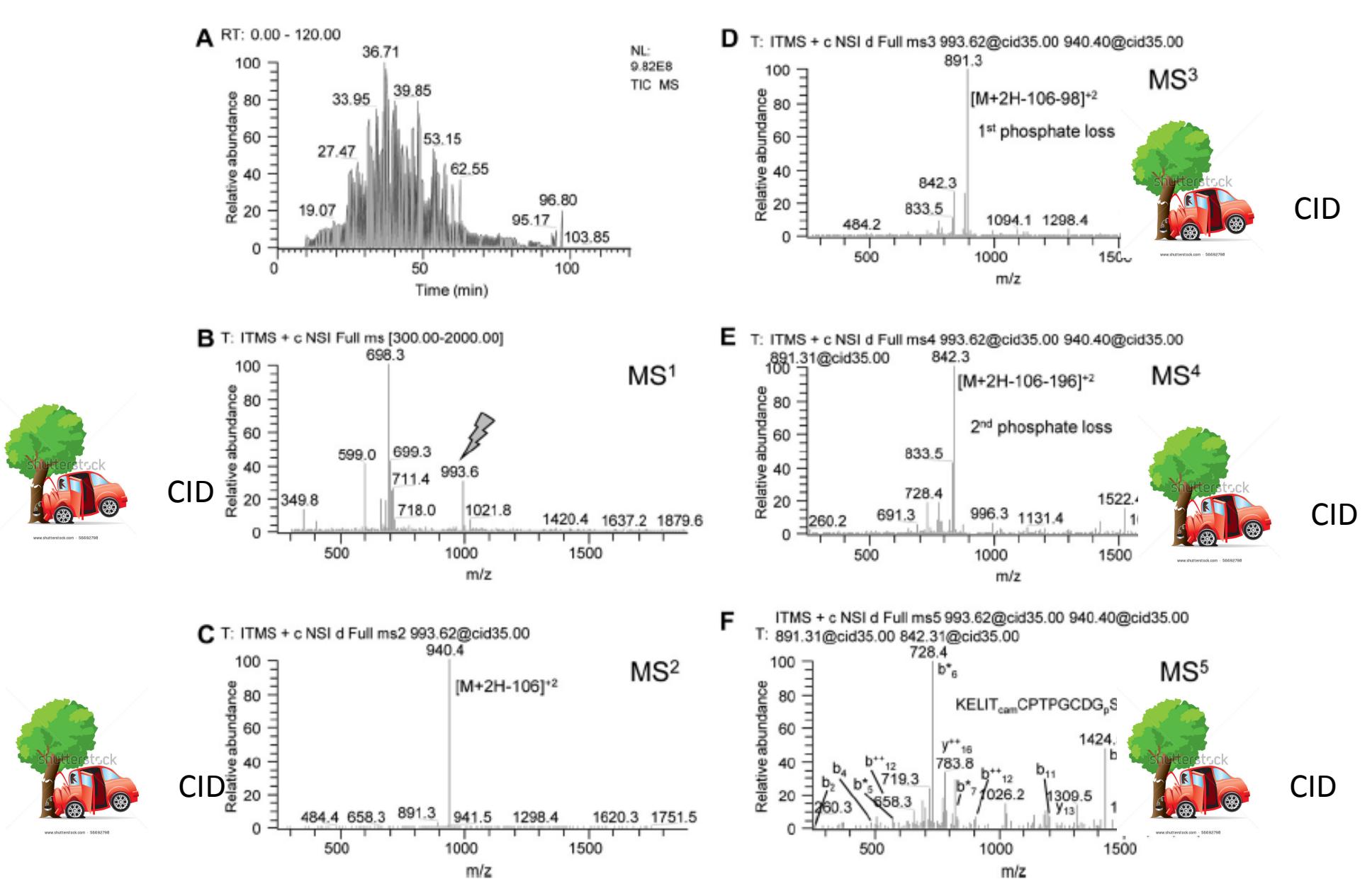
Collision induced dissociation CID

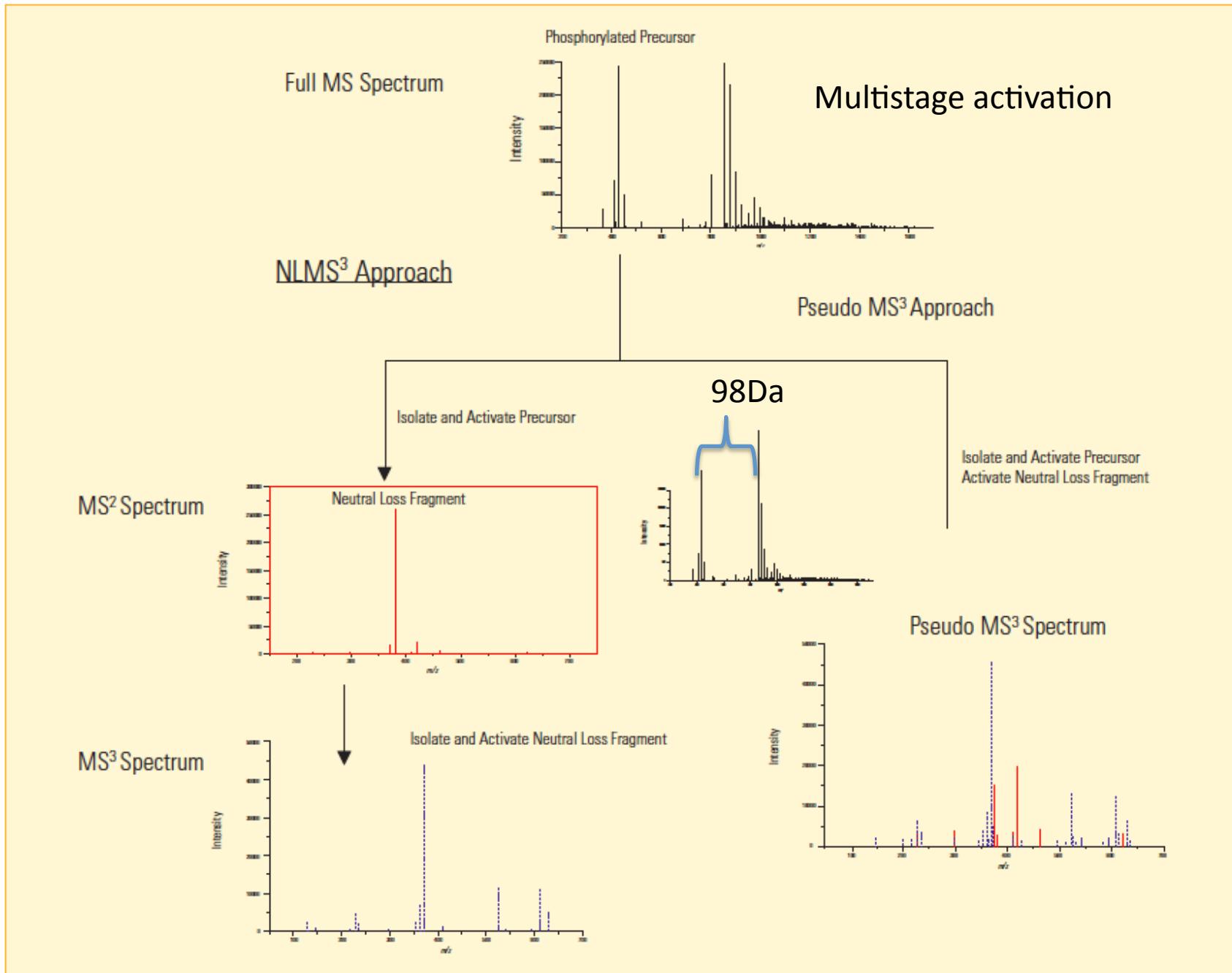


www.shutterstock.com · 56692798

MS/MS using CID usually generates the dominant neutral loss of H₃PO₄ (-97.98Da) from Serine and threonine, absorbs energy

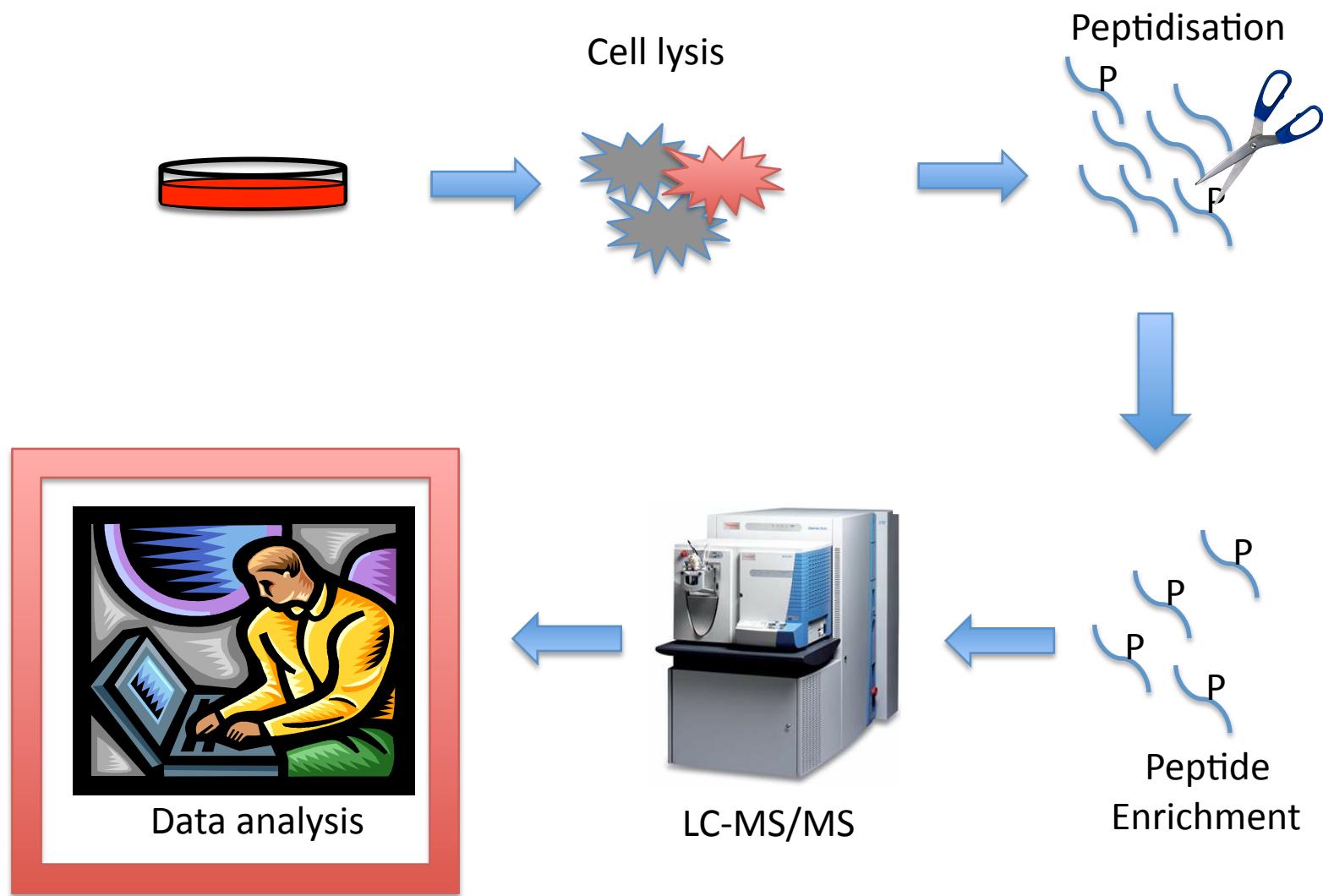




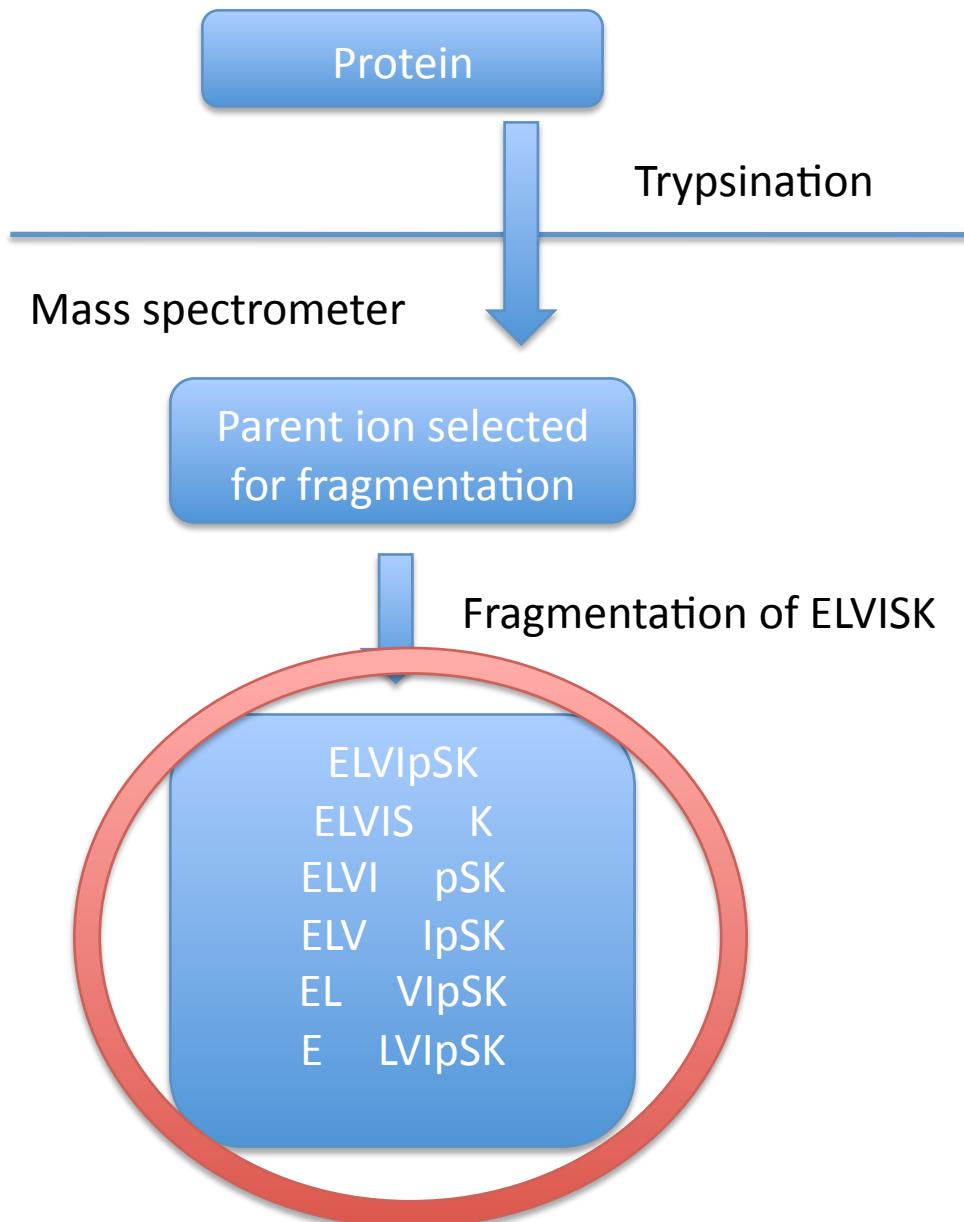


Thermo Scientific

Global phosphoproteomics approach



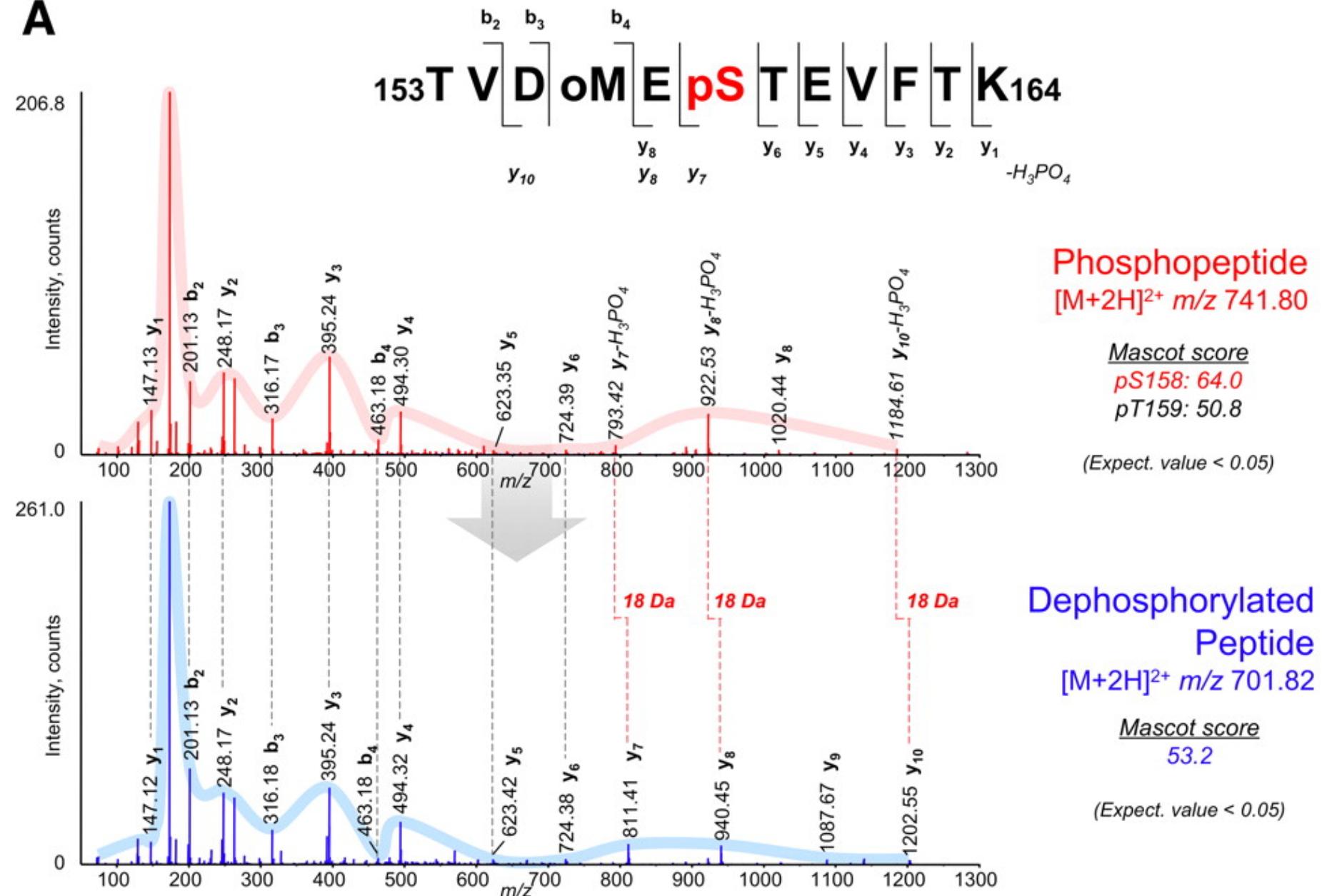
Wet lab



Mapping of PTMs with MS

Does the PTM remain stable during MS/MS fragmentation?					Is the PTM labile during MS/MS fragmentation?
Look for characteristic mass shifts in parent and fragment ions:		Look for MS/MS signature ions formed during fragmentation:			Monitor products of labile cleavage (such as neutral loss):
Phosphorylation	+80 Da	pY pS, pT, pY	216 Da -79, -63 Da		-98 Da



A

Wet lab

Protein

Trypsination

Mass spectrometer

Parent ion selected
for fragmentation

Fragmentation of ELVISK

ELVISK

ELVIS K

ELVI SK

ELV ISK

EL VISK

E LVISK

Computer

Database

Trypsination
in silico

Wet lab

Protein

Trypsination

Mass spectrometer

Parent ion selected
for fragmentation

Fragmentation of ELVISK

ELVISK
ELVIS K
ELVI SK
ELV ISK
EL VISK
E LVISK

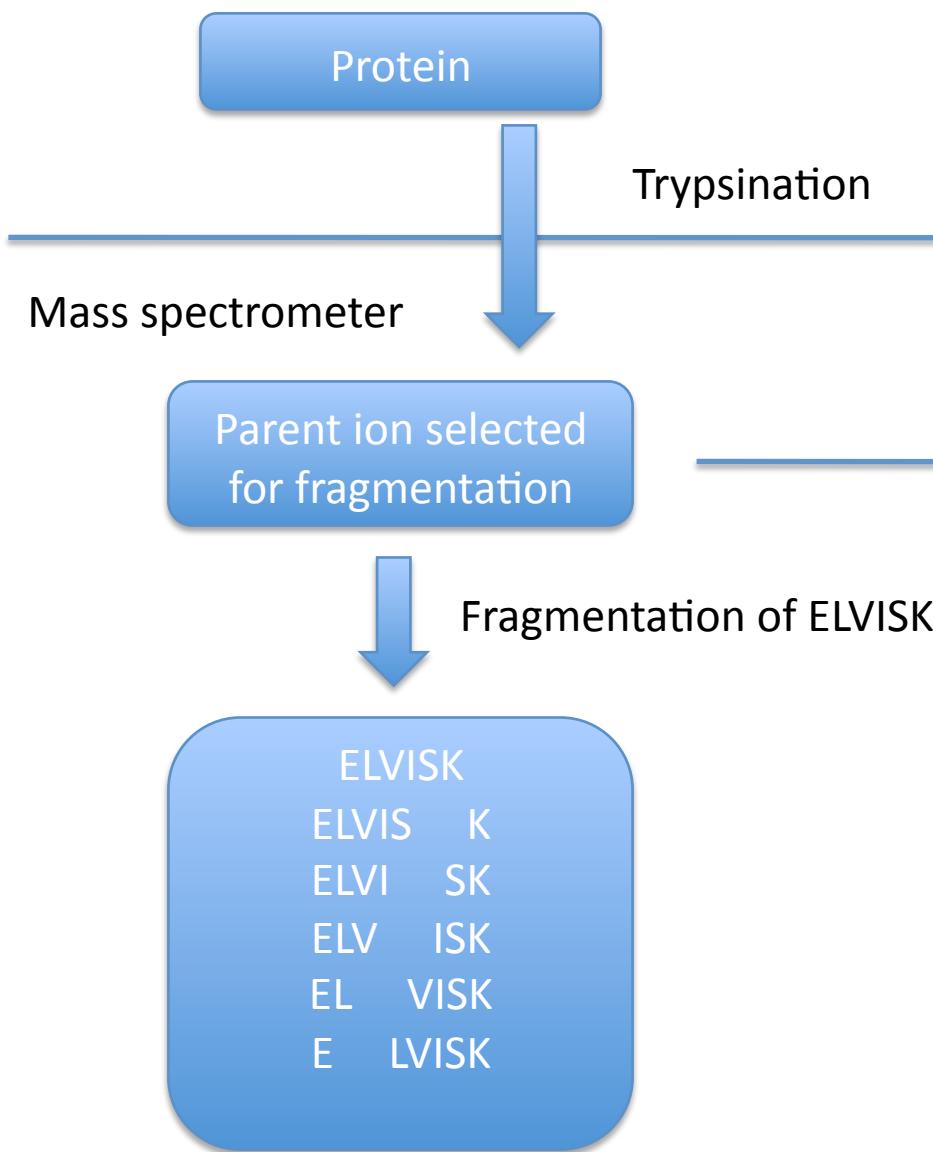
Computer

Database

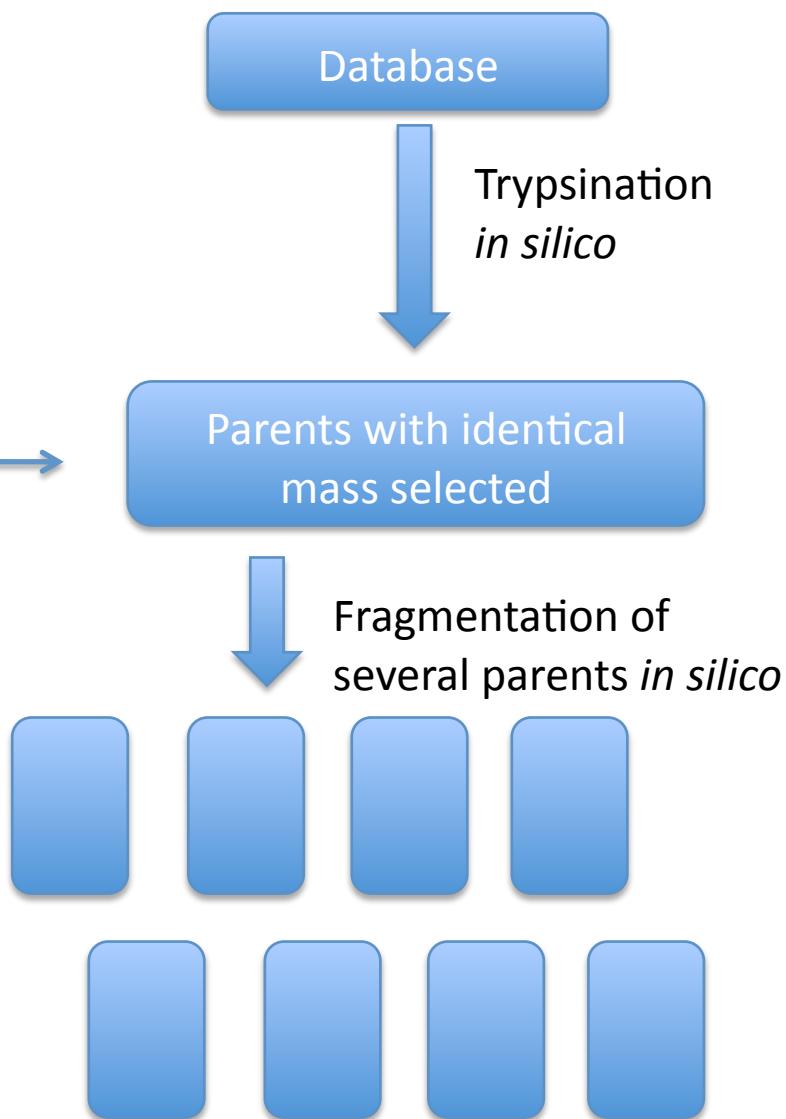
Trypsination
in silico

Parents with identical
mass selected

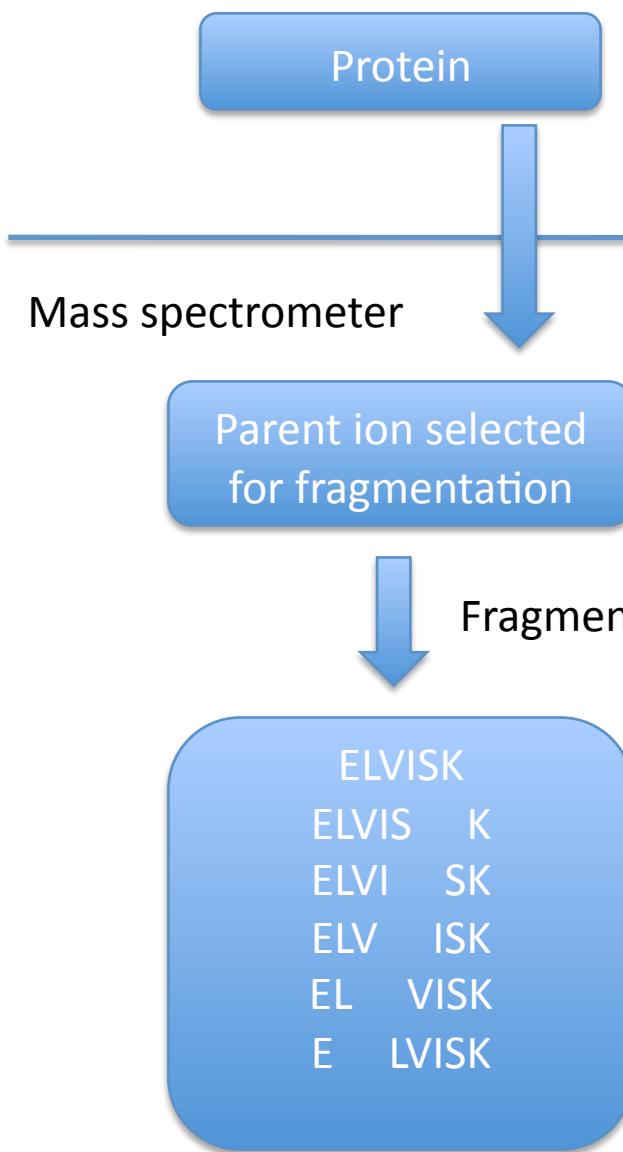
Wet lab



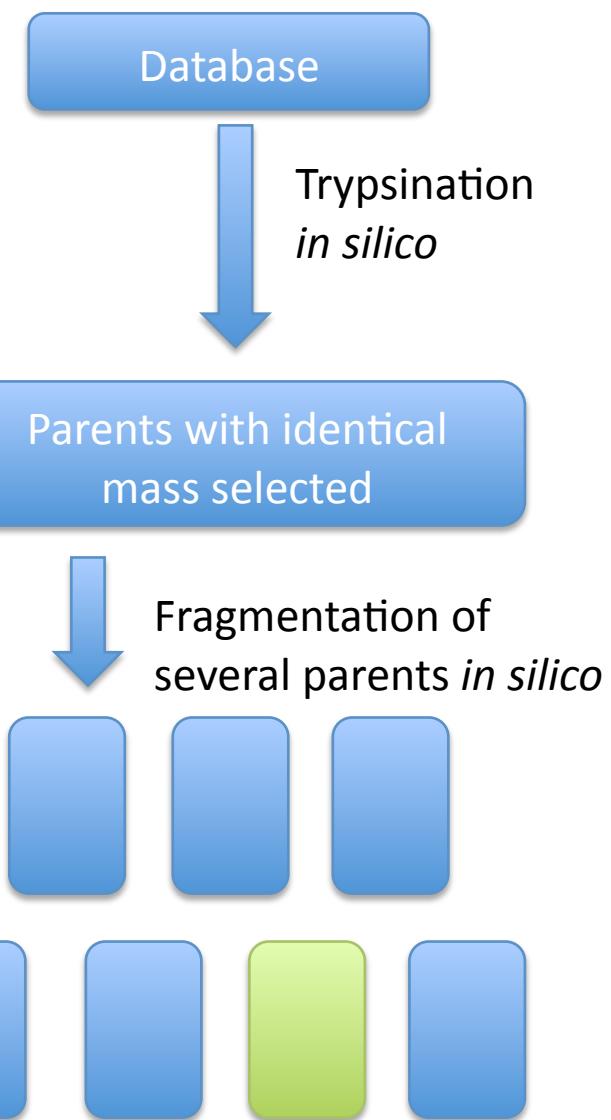
Computer



Wet lab



Computer



Why to map phosphorylation sites

How to map phosphorylation sites

